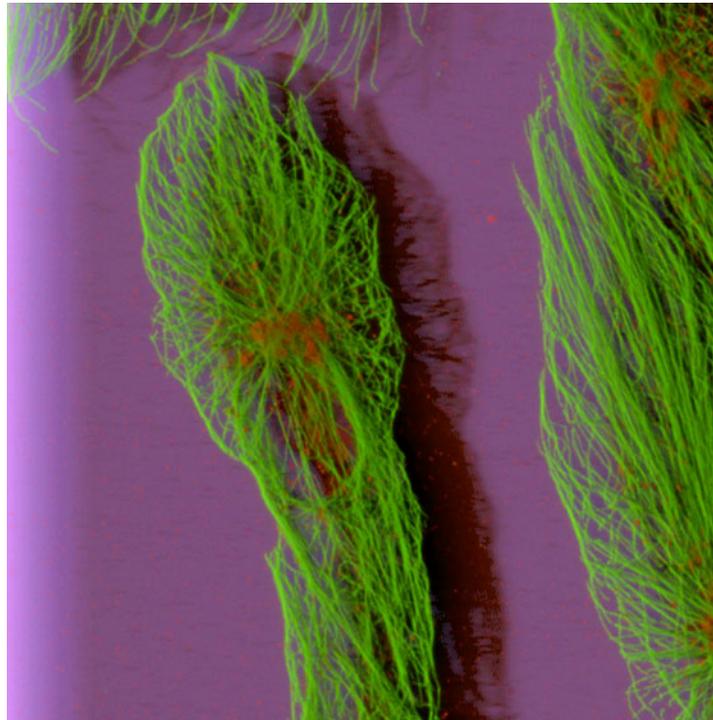


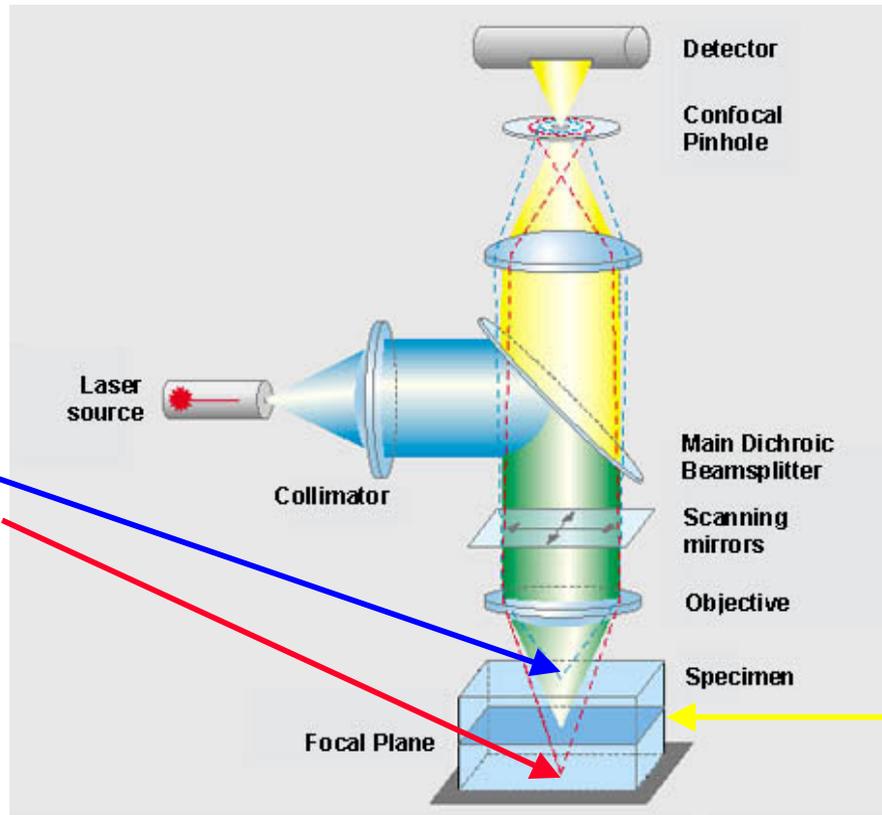
# Confocal microscopy

Zeiss LSM 510 and Zeiss LSM 510 META

Visualisation of biological structures in 3D



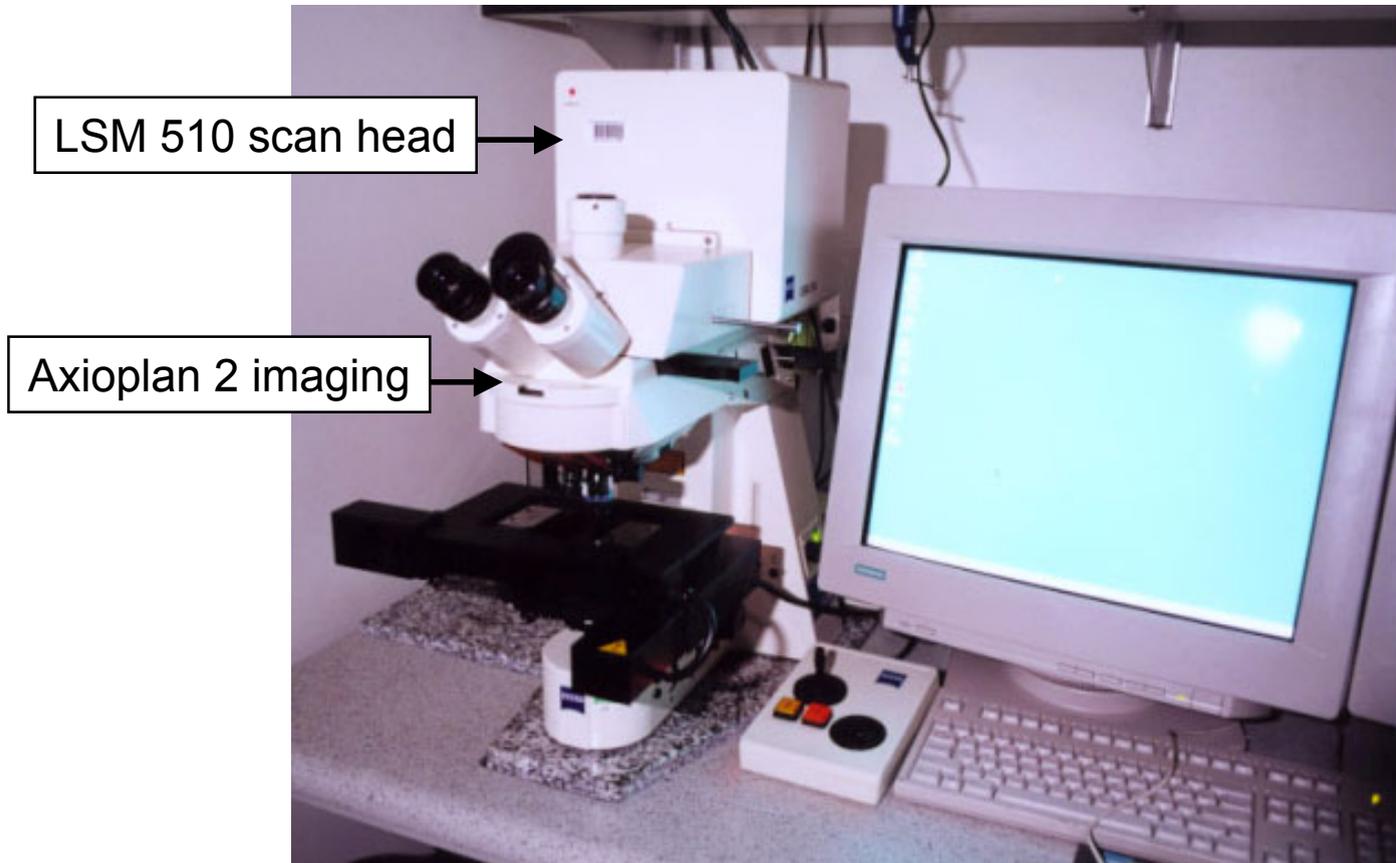
## Confocal Principle



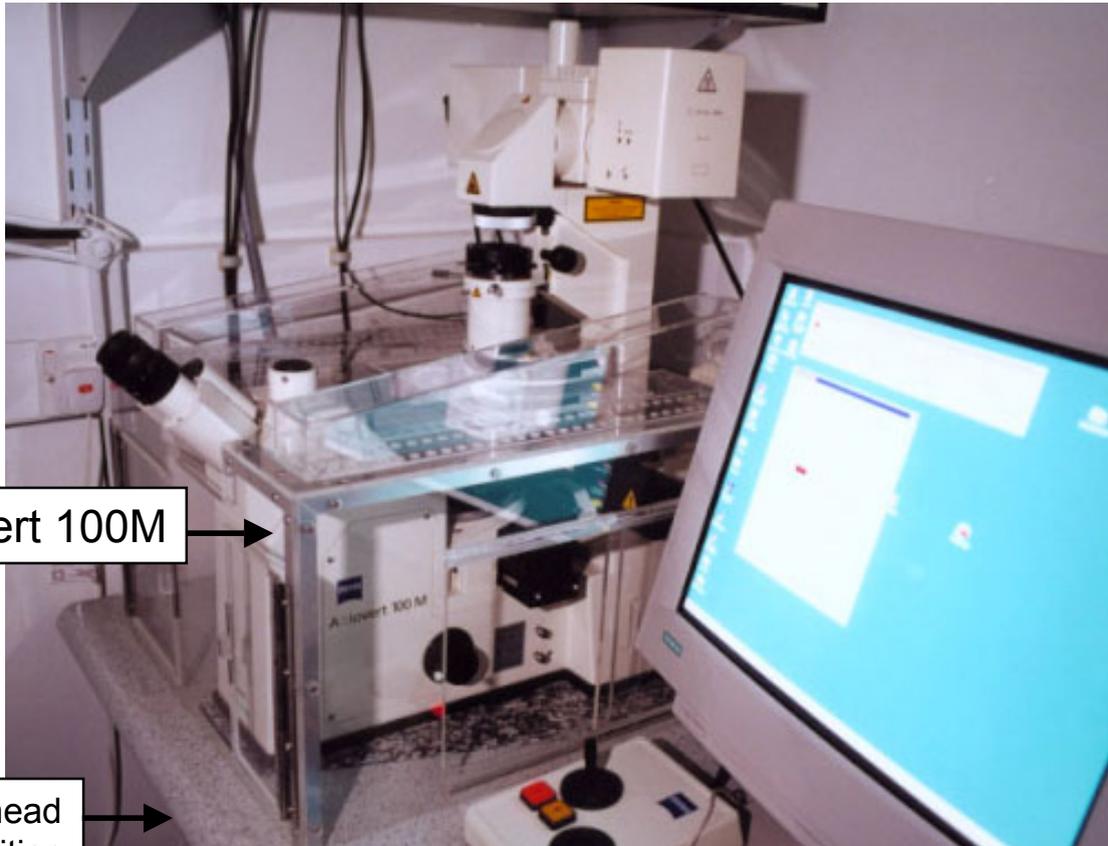
Signals from above and below the plane of focus fall outside the pinhole and are blocked

Only signals from plane of focus pass the pinhole and are detected - producing a "optical section"

## Upright Zeiss LSM 510 confocal microscope



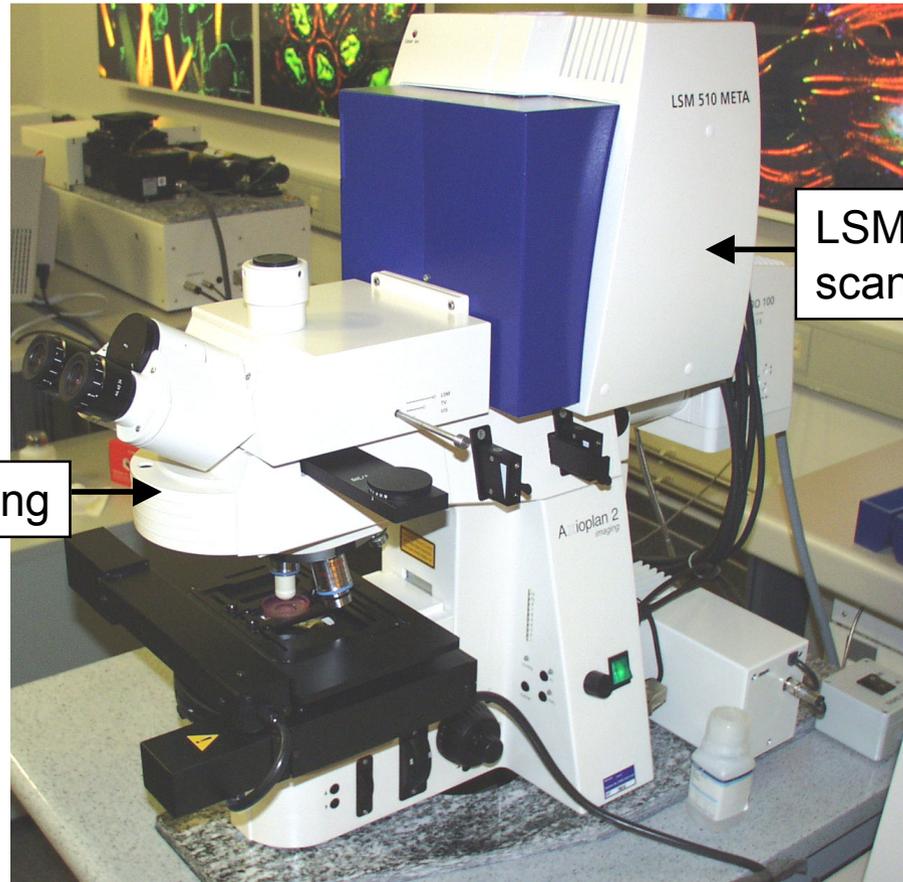
## Inverted Zeiss LSM 510 confocal microscope



Axiovert 100M

LSM 510 scan head  
in base port position

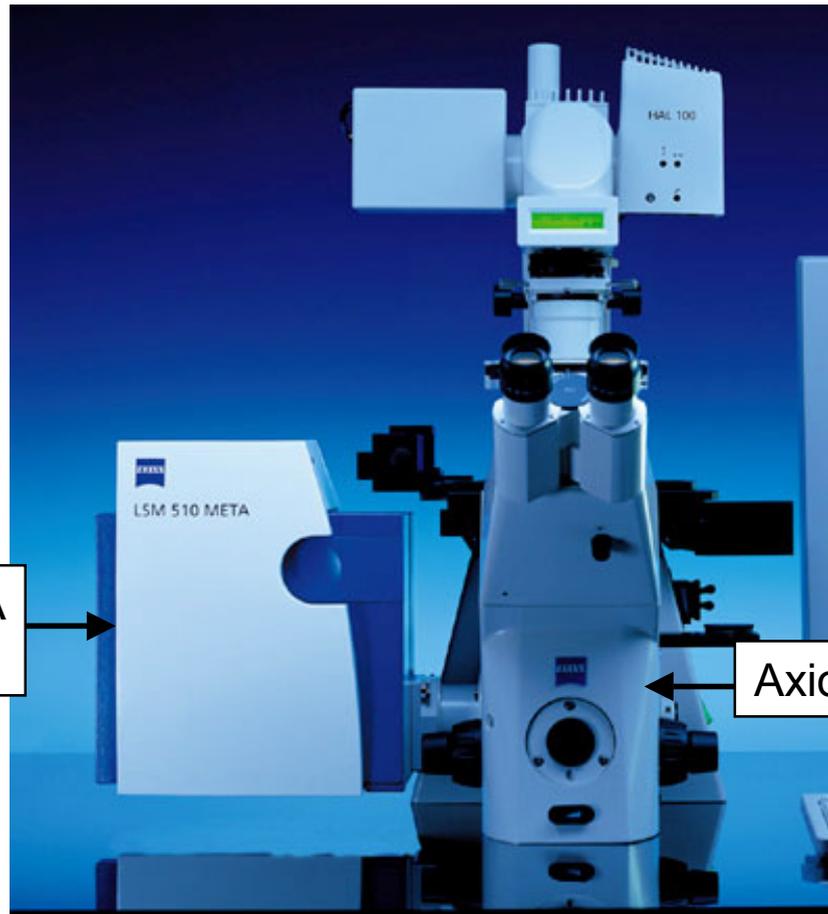
## Upright Zeiss LSM 510 META confocal microscope



Axioplan 2 imaging

LSM 510 META  
scan head

## Inverted Zeiss LSM 510 META confocal microscope



LSM 510 META  
scan head

Axiovert 200M



## Contents

- Starting the Zeiss LSM 510 microscope, software and laser  
Selecting an objective and focusing the microscope
- Selecting an objective and focusing the microscope
- Configuring the laser scanning and detection for confocal image acquisition
- Acquiring a Z- and Time - Series
- Data storage

Descriptions also include the LSM 510 META



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- Starting the Zeiss LSM 510 microscope, software and laser  
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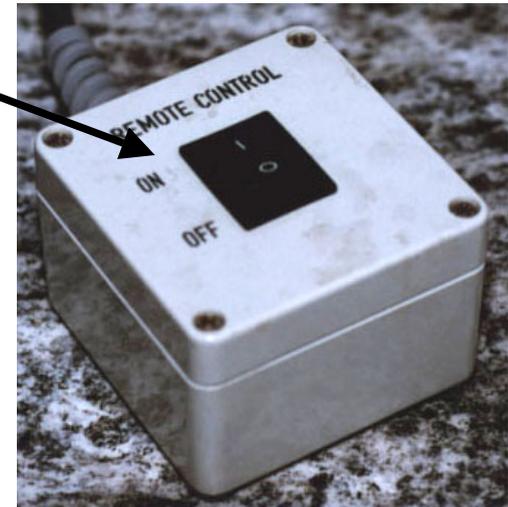
Descriptions also include the LSM 510 META

## Start the Zeiss LSM 510 Confocal Microscope

1) First switch on the mercury lamp



2) Turn on the remote control switch



3) Wait for the computer to boot up and Login by simultaneously pressing the Ctrl, Alt and Delete keys

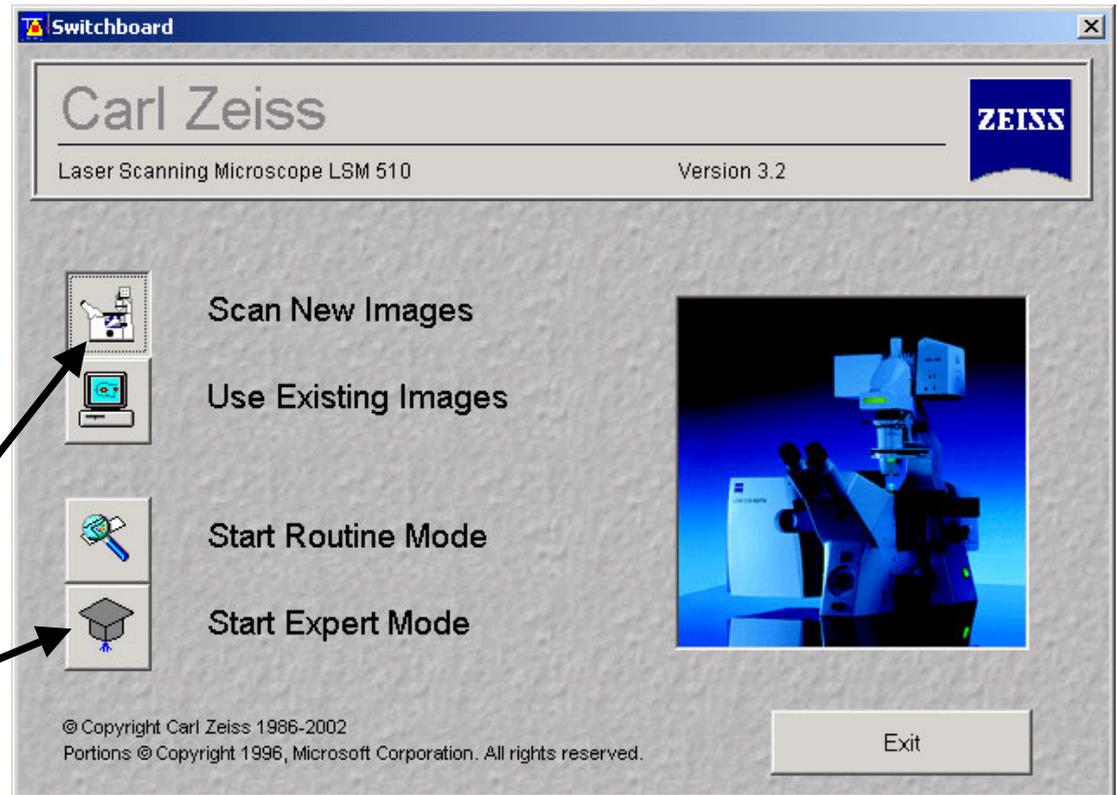
## Starting the LSM 510 software



1) Double click the  
LSM 510 icon

2) Select "Scan New  
Images"

3) Select "Start  
Expert Mode"

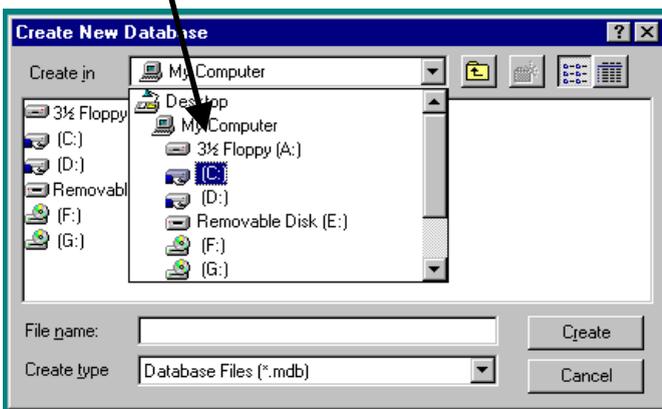


## Creating a database for acquired images



1) In the main menu *File* select *New* database

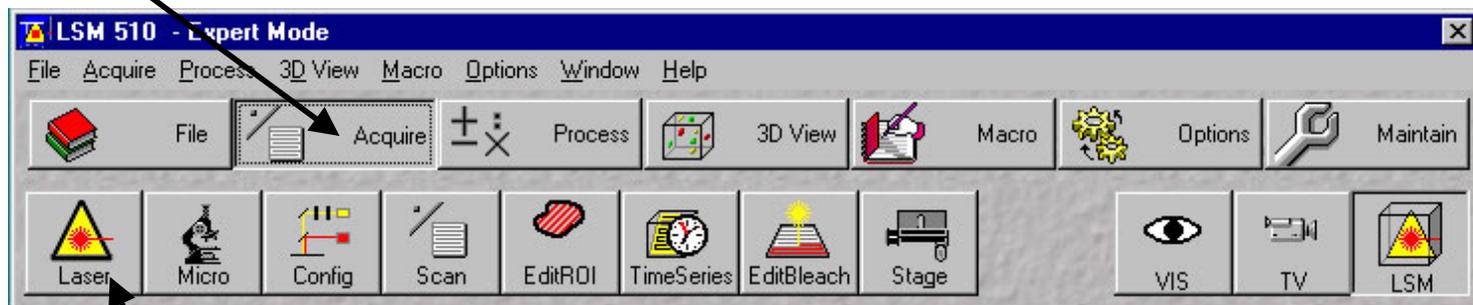
2) Select drive C or D: from pull down menu



3) Create a new directory if needed



## 1) Select Acquire Turning on the lasers

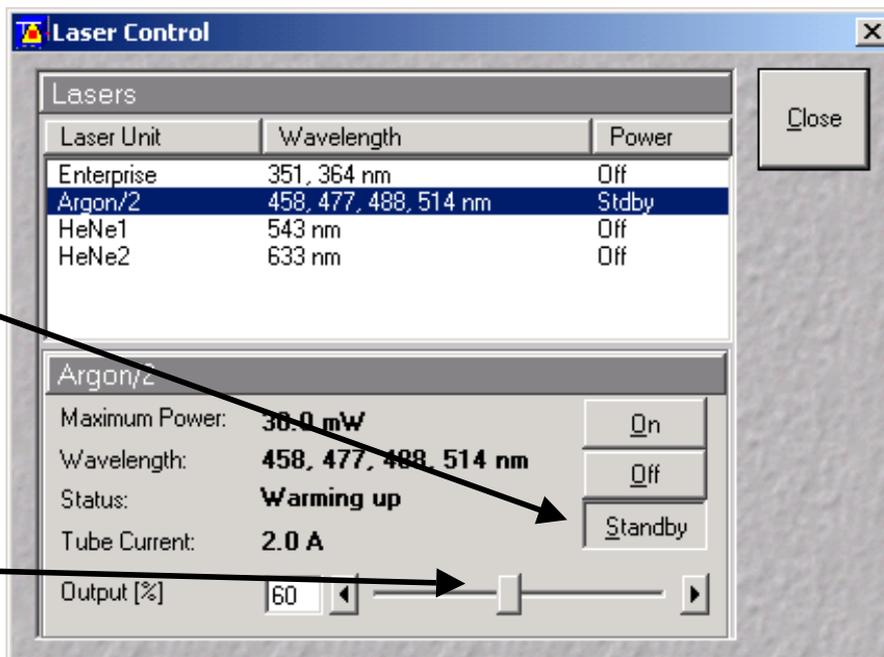


2) Select *Laser*

3) Switch required laser/s to *Standby*

4) When status is *Ready*, click *On*

5) Set Output [%] so that the tube current is between 5.5 and 6.5 A

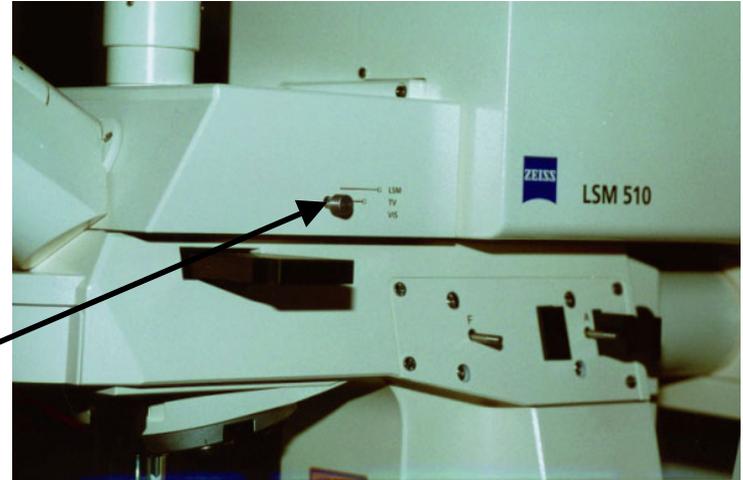


## Change between direct observation and laser scanning

### Upright Microscopes: Axioplan 2 imaging and Axioskop 2 FS

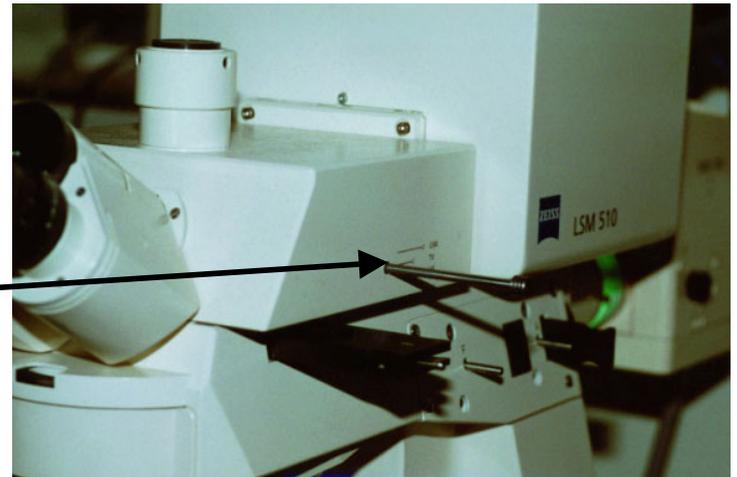
For direct observation of transmitted light and fluorescence:

Set slider to “VIS” (push it in)



For laser scanning image acquisition:

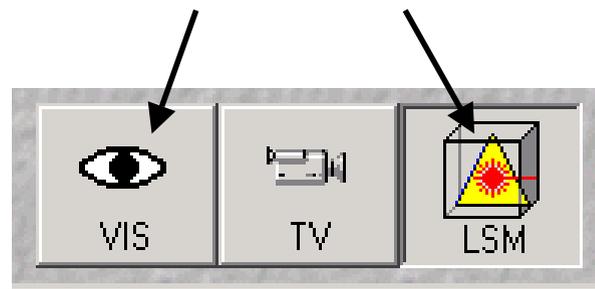
Set slider to “LSM”  
(pull slider out)



## Change between direct observation and laser scanning

### Inverted Microscope: Axiovert 200 M

Toggle between Vis and LSM button in main menu, automatic switching between direct observation and laser scanning (no slider)





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Descriptions also include the LSM 510 META

## Selecting an objective and focusing the microscope

The screenshot shows the 'LSM 510 - Expert Mode' main window and the 'Microscope Control' sub-window. In the main window, the 'Acquire' menu is open, and the 'Micro' button is highlighted with an arrow. In the 'Microscope Control' window, the 'Objective' section is expanded, showing a list of objective lenses. The 'Plan-Apochromat 63x/1.4 Oil' objective is selected and highlighted in blue. The 'Transmitted Light' and 'Reflected Light' sections are also visible, showing their respective settings.

1) Select *Micro* (Main menu: *Acquire*)  
(For Axioskop 2 FS these settings have to be adjusted manually)

2) Microscope settings can be stored and up to 8 buttons assigned for fast retrieval and adjustment

3) Objective lens can be selected from a pull down menu by clicking onto the *Objective* button

## Focusing the microscope in fluorescence mode

The screenshot displays the 'LSM 510 - Expert Mode' software interface. The top menu bar includes 'File', 'Acquire', 'Process', '3D View', 'Macro', 'Options', 'Maintain', 'Window', and 'Edit'. Below the menu is a toolbar with icons for 'File', 'Acquire', 'Process', and '3D View'. A secondary toolbar contains icons for 'Laser', 'Micro', 'Config', 'Scan', 'Edit ROI', 'TimeSeries', and 'Edit'.

The 'Microscope Control' window is open, showing 'Microscope Settings' with a dropdown menu set to 'DAPI'. Below this are buttons for 'Apply', 'Store', 'Delete', and 'Assign Button'. A table displays filter settings:

DAPI	FITC	TRITC	n.n
n.n	n.n	n.n	n.n

The main control area features a light path diagram. On the right, 'Transmitted Light' is shown as 'Off 0.0%' with a crossed-circle icon. Below it, 'Reflected Light' is shown with a crossed-circle icon. The 'Objective' is identified as 'Achromplan IR 40x/0.8 W'. A 'Reflector' control is shown with a pull-down menu containing options: 'None', 'DAPI', 'GFP', 'Rhod', and 'None'. The 'GFP' option is currently selected. Arrows from the text below point to the 'Reflected Light' icon and the 'Reflector' menu.

Click onto *Reflected Light* to open the shutter of the HBO lamp

Fluorescence is observed by selecting the appropriate filter set in the pull down menu of *Reflector*

## Focusing the microscope in transmitted mode

Click onto *Transmitted Light* and move the slider to set the intensity of the HAL illumination

Use no reflector cube in the reflector turret, chose *None*

**Microscope Control**

**Microscope Settings**

DAPI [Apply] [Store] [Delete] [Assign Button]

DAPI	FITC	TRITC	n.n
n.n	n.n	n.n	n.n

**Transmitted Light**

[Close]

[Less]

[Close]

On 3200 K

None

**Tube Lens**  
Lens LSM

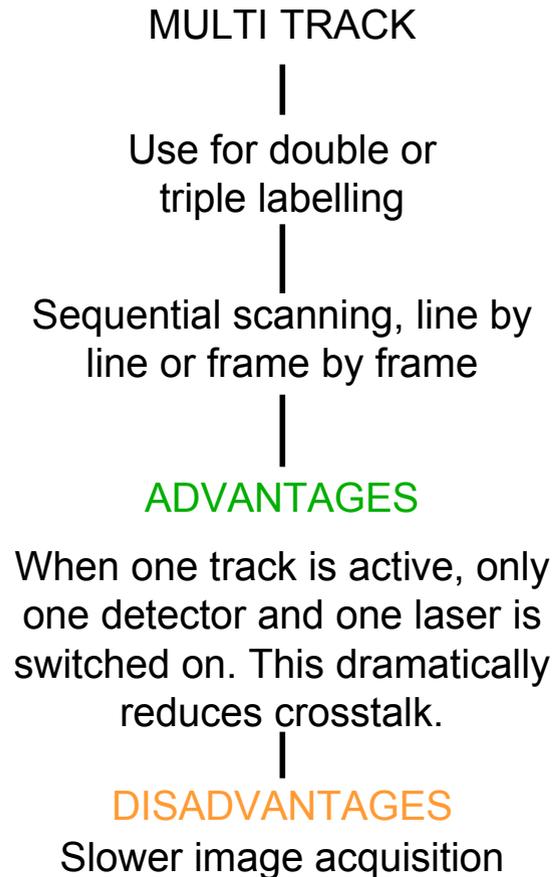
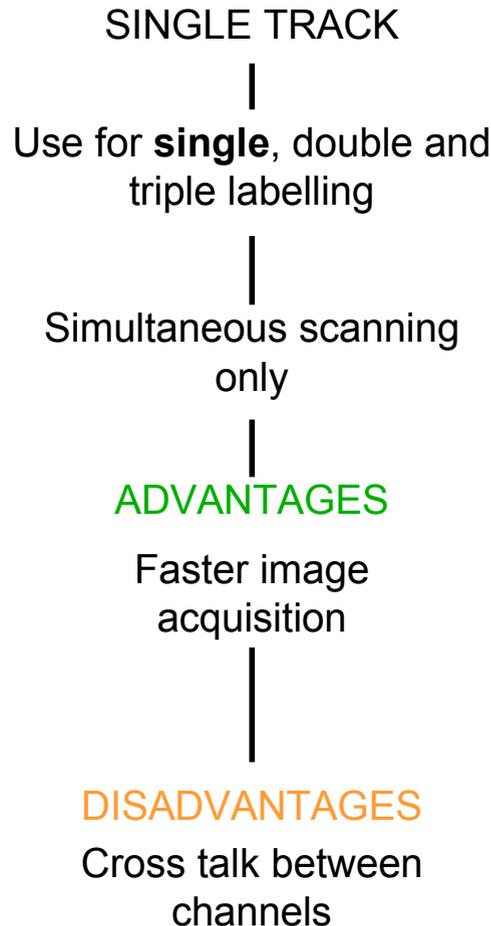


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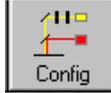
Descriptions also include the LSM 510 META

## Choosing the configuration



## Configuration of the filters and storage of the track configurations

SINGLE TRACK - lasers scan simultaneously

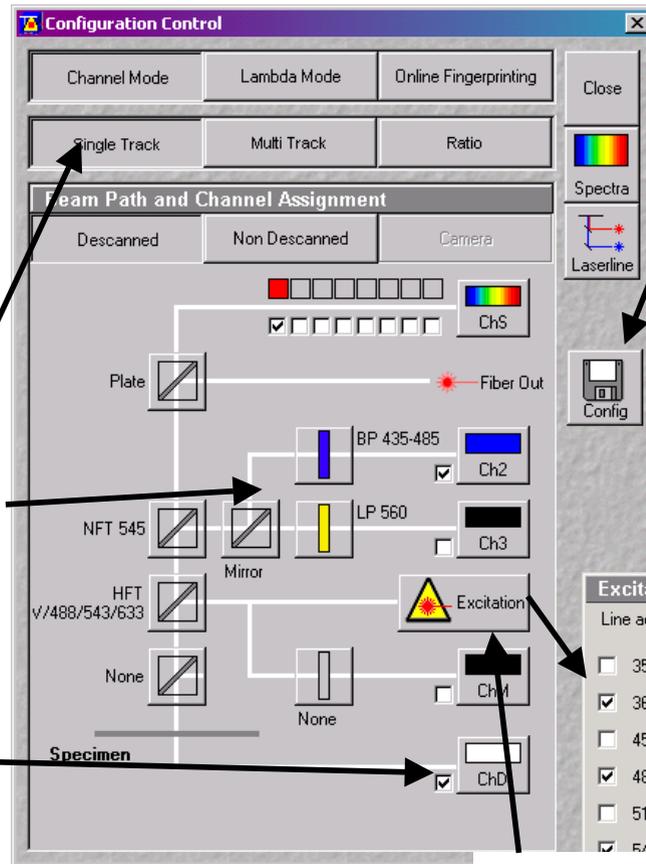


1) Select *Config* in the *Acquire* menu

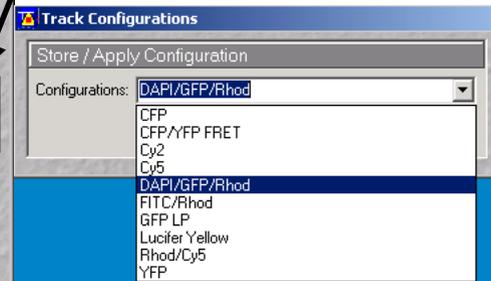
2) Select *Single Track*

4a) Select the appropriate filters and activate the Channels

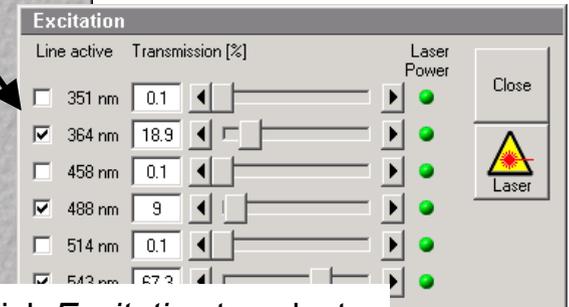
4c) Transmitted light image can also be generated. Transmission channel is usually set to white colour.



4b) The *Config* button opens the pull down menu to load/store Track configurations



3) Click *Excitation* to select the laser and attenuation



## Applying a stored configuration and checking the settings



If you select *Store* by mistake, software will ask you, if you want to overwrite the configuration. **ANSWER NO!**

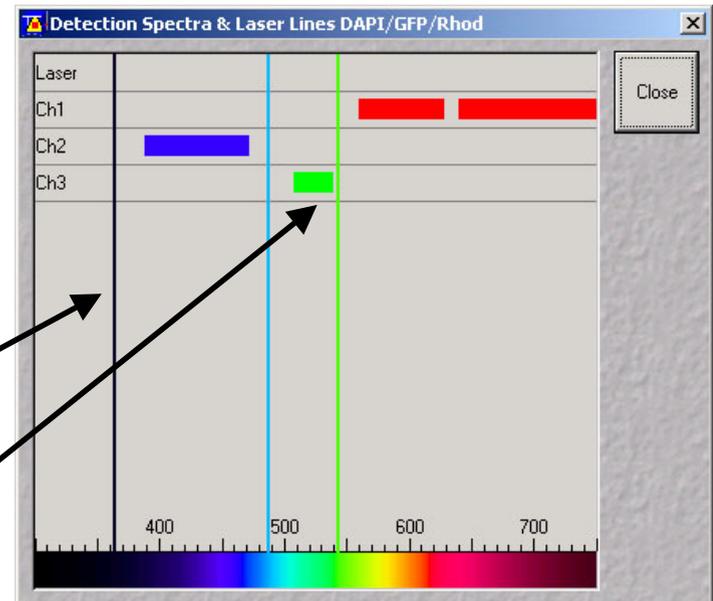
Each new login loads a predefined set of correct configurations.

5) Chose a configuration in the *Track Configuration* menu.  
Select *Apply*

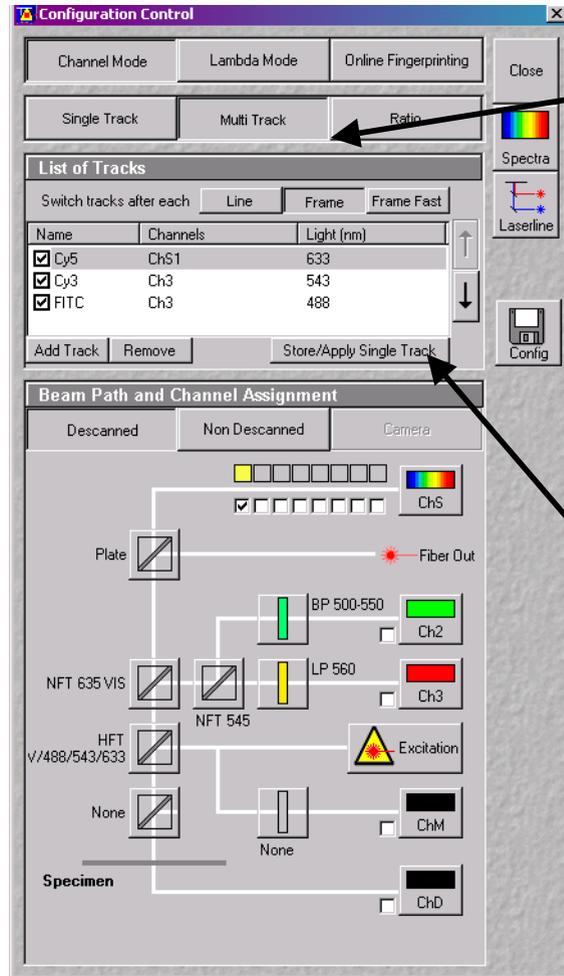
6) To check for correct settings, click the *Spectra* button



The *Spectra* button opens a window to display the activated laser lines for excitation (colored vertical lines) and channels (colored horizontal bars)



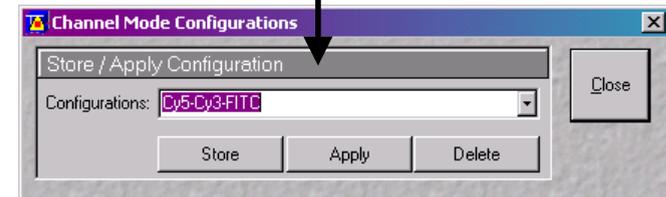
## Multi Track Configuration



1) Select *Multi Track* for sequential scanning

2) Select *Config*

3) Select a stored track from the pull down menu, click on *Apply*



This button stores only the highlighted single track or applies a single track.

## Cy5-Cy3-FITC Multi Track

Three laser lines and channels activated sequentially

### Excitation

Excitation control panel showing 633 nm selected. The table below summarizes the active state:

Line active	Transmission [%]	Laser Power
<input type="checkbox"/> 458 nm	0.1	Off
<input type="checkbox"/> 488 nm	16.9	Off
<input type="checkbox"/> 514 nm	0.1	Off
<input type="checkbox"/> 543 nm	0.1	Off
<input checked="" type="checkbox"/> 633 nm	61.4	On
<input type="checkbox"/> 880 nm	0.1	Off

633 nm, using the META detector in Channel mode

Excitation control panel showing 543 nm selected. The table below summarizes the active state:

Line active	Transmission [%]	Laser Power
<input type="checkbox"/> 458 nm	0.1	Off
<input type="checkbox"/> 488 nm	0.1	Off
<input type="checkbox"/> 514 nm	0.1	Off
<input checked="" type="checkbox"/> 543 nm	43.6	On
<input type="checkbox"/> 633 nm	0.1	Off
<input type="checkbox"/> 880 nm	0.1	Off

543 nm

Excitation control panel showing 488 nm selected. The table below summarizes the active state:

Line active	Transmission [%]	Laser Power
<input type="checkbox"/> 458 nm	0.1	Off
<input checked="" type="checkbox"/> 488 nm	3	On
<input type="checkbox"/> 514 nm	0.1	Off
<input type="checkbox"/> 543 nm	0.1	Off
<input type="checkbox"/> 633 nm	0.1	Off
<input type="checkbox"/> 880 nm	0.1	Off

488 nm

### Detection

Detection setup for 633 nm. The META detector is selected in Channel mode. The table below summarizes the active state:

Component	State
Plate	Active
BP 500-550	Off
Ch2	Off
LP 560	Off
Ch3	Off
Excitation	On
ChM	Off

Detection setup for 543 nm. The META detector is selected in Channel mode. The table below summarizes the active state:

Component	State
Plate	Active
BP 500-550	Off
Ch2	Off
LP 560	Off
Ch3	On
Excitation	On
ChM	Off

Detection setup for 488 nm. The META detector is selected in Channel mode. The table below summarizes the active state:

Component	State
Plate	Active
BP 500-550	On
Ch2	On
LP 560	Off
Ch3	Off
Excitation	On
ChM	Off

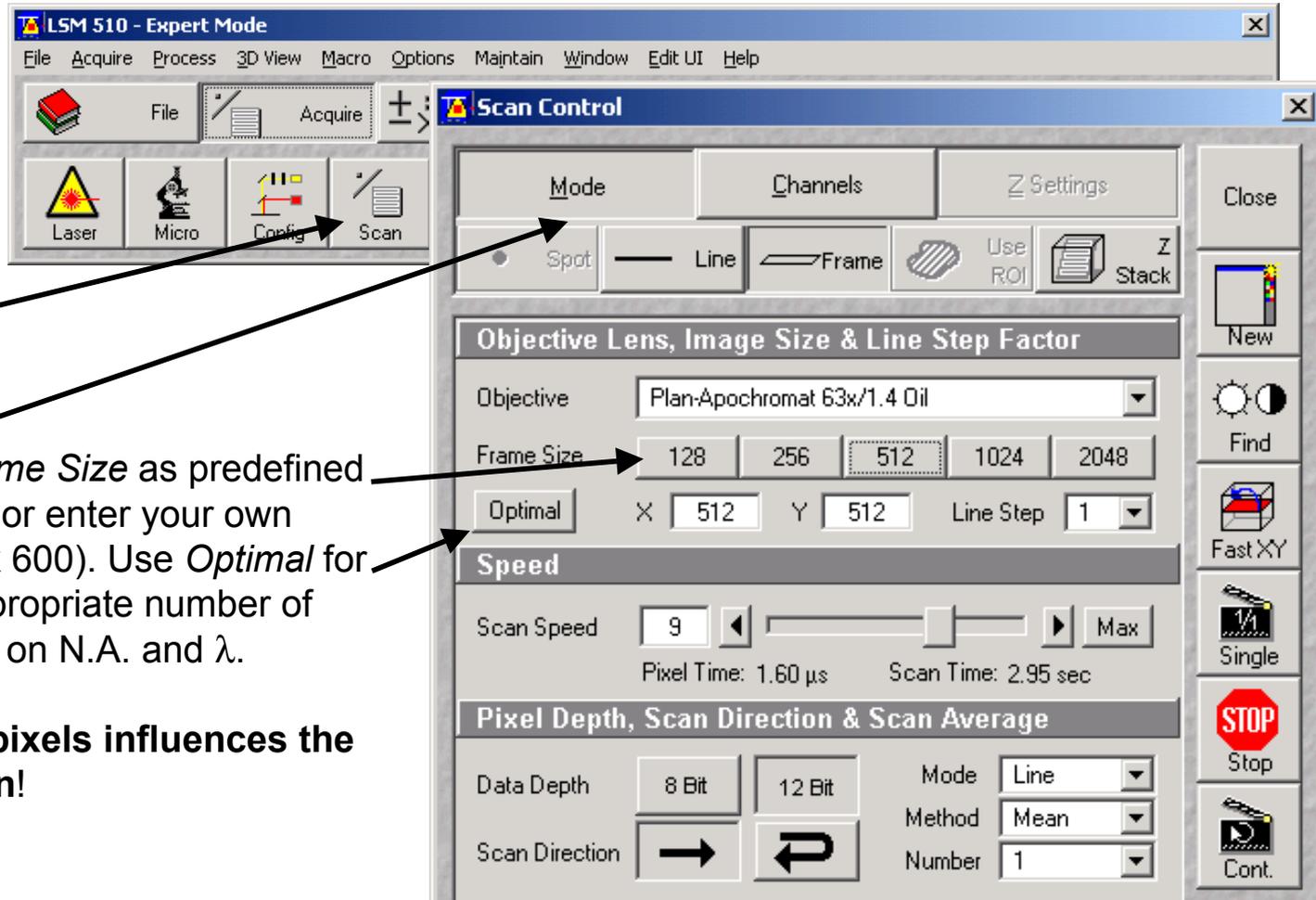
## Setting the parameters for scanning

1) Select *Scan*

2) Select *Mode*

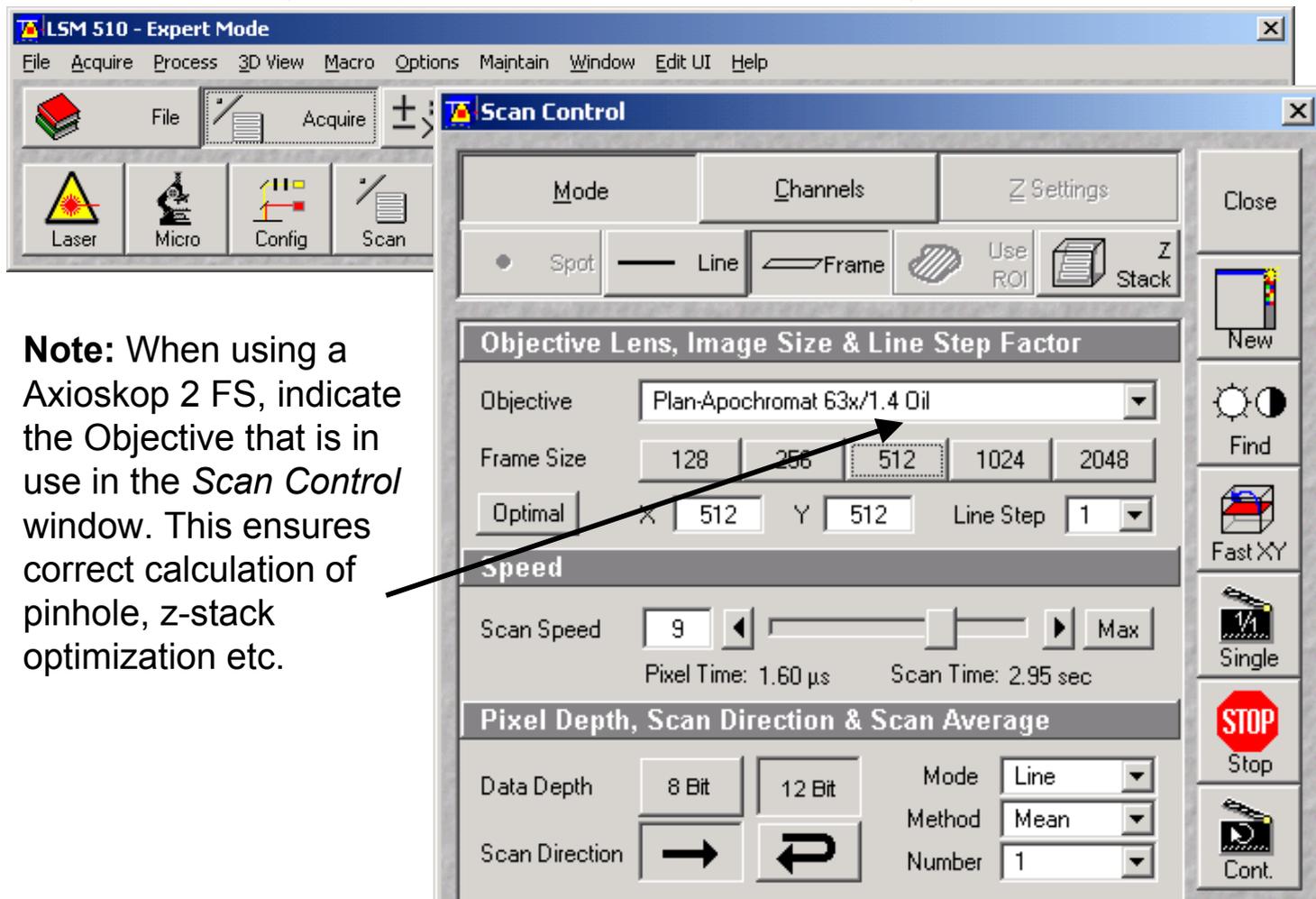
3) Select the *Frame Size* as predefined number of pixels or enter your own values (e.g 300 x 600). Use *Optimal* for calculation of appropriate number of pixels depending on N.A. and  $\lambda$ .

**The number of pixels influences the image resolution!**

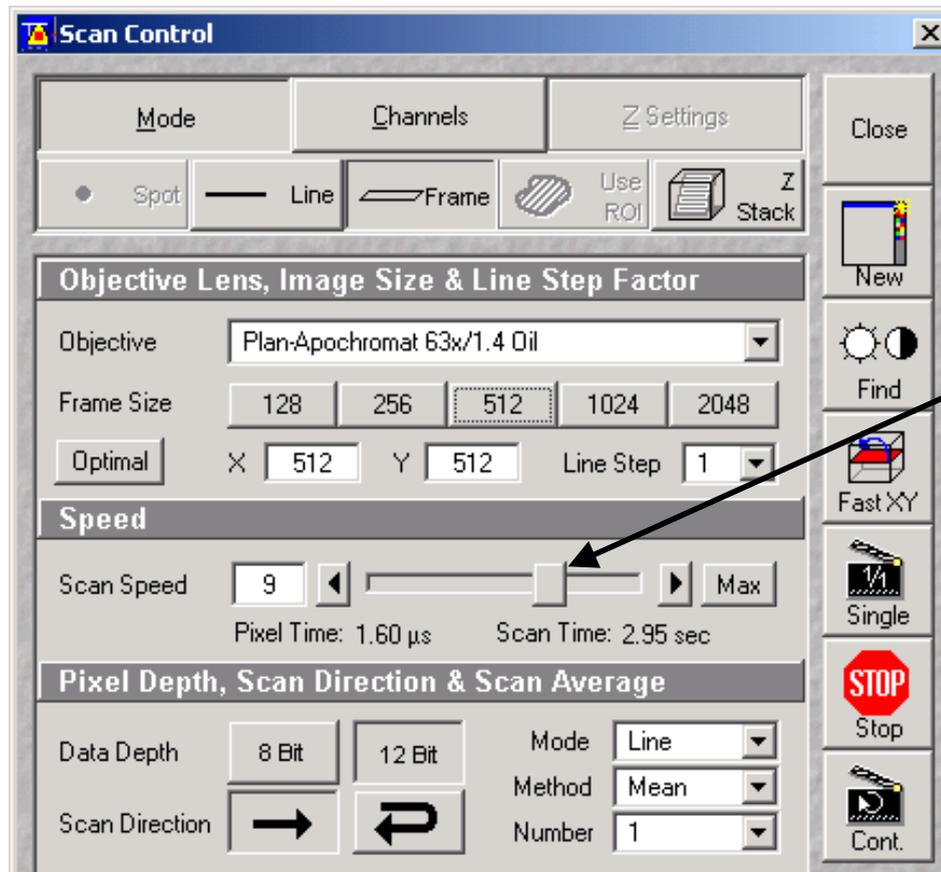


## Setting the parameters for scanning

**Note:** When using a Axioskop 2 FS, indicate the Objective that is in use in the *Scan Control* window. This ensures correct calculation of pinhole, z-stack optimization etc.

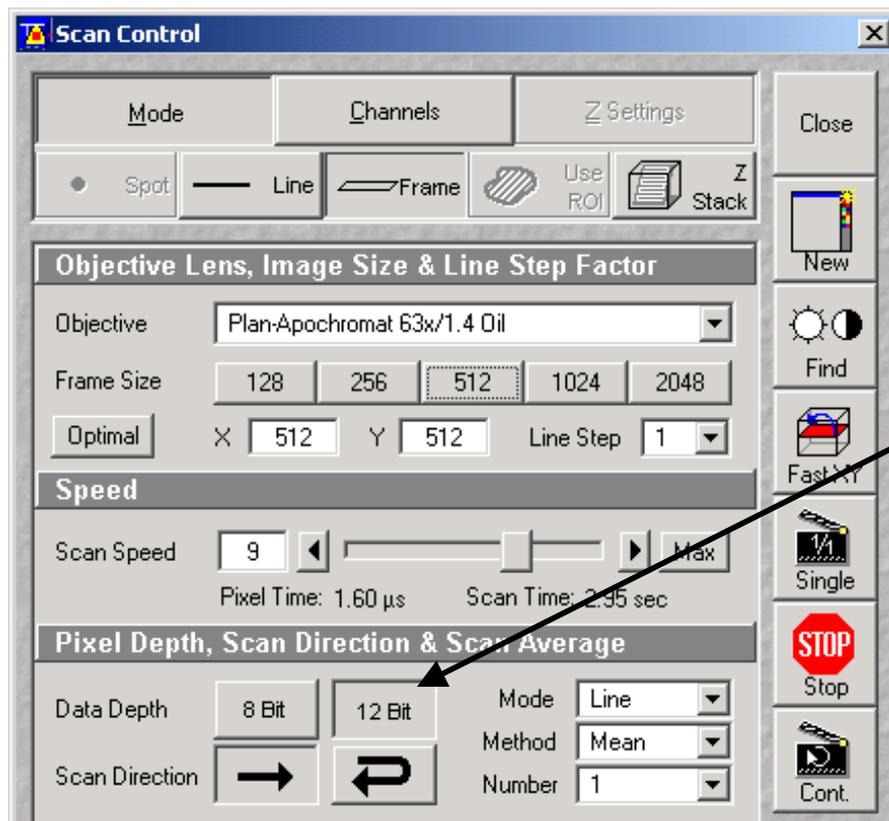


## Adjusting the scan speed



Adjust the scan speed - a higher speed with averaging results in the best signal to noise ratio. Scan speed 8 usually produces good results. Use 6 or 7 for superior images.

## Choosing the Dynamic Range (8/12 Bit per pixel)

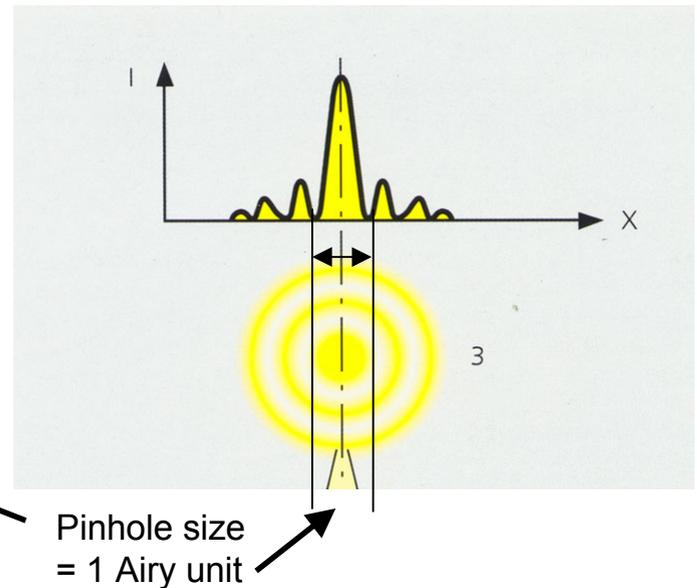
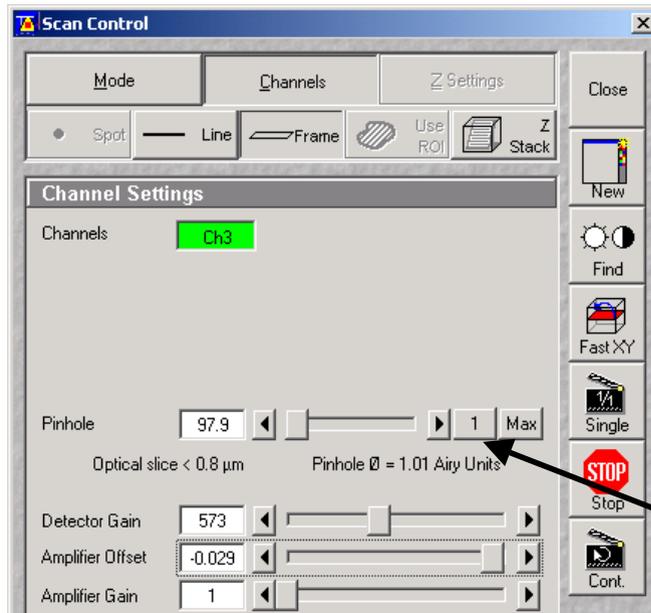


Select the dynamic range - 8 bit will give 256 gray levels, 12 Bit will give 4096 levels. Photoshop 5 will import 12 and 16 Bit images.

Publication quality images should be acquired using 12 Bit.

12 Bit is also recommended when doing quantitative measurements or when imaging low fluorescence intensities.

## Channel Settings - Adjusting the Pinhole



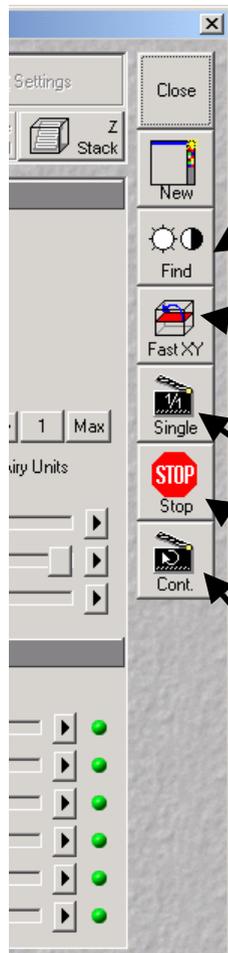
Set pinhole size to 1 Airy unit for best compromise between depth discrimination and efficiency.

Pinhole adjustment changes the “Optical slice”.

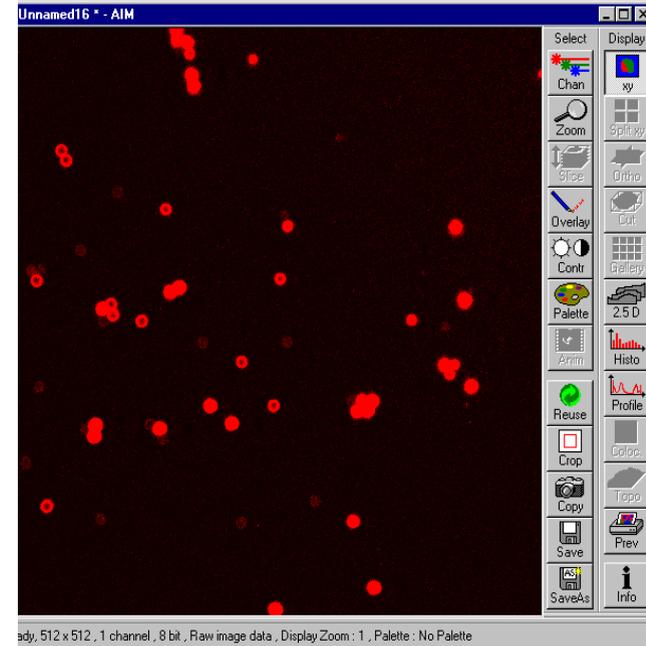
*When collecting multi channel images, adjust the pinholes so that each channel has the same “Optical Slice”.*

This is important for colocalization studies.

## Image Acquisition

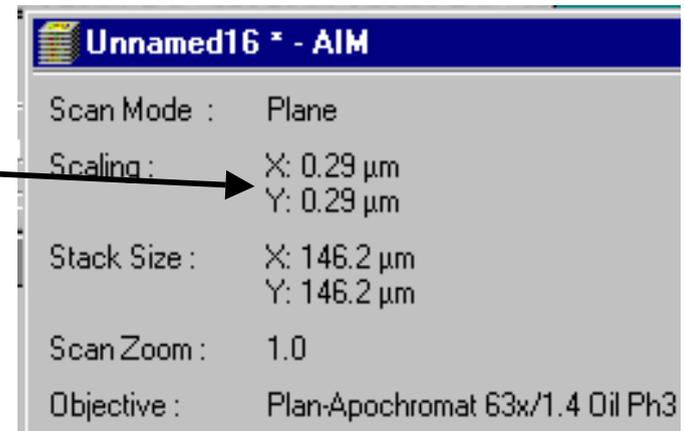


- 1) *Find* opens new image window and automatically pre-adjusts detector sensitivity
- 2) Select *Fast XY* for continuous fast scanning - useful for finding and changing the focus
- 3) *Single* records a single image
- 4) *Stop* blanks the laser beam and stops the scanning mirrors
- 5) Select *Cont.* for continuous scanning with selected scan speed



## Minimal Pixel Size determined by Nyquist Sampling

Magnification	NA	Pixel size
5	0.15	1.03 $\mu\text{m}$
10	0.3	0.51 $\mu\text{m}$
20	0.5	0.31 $\mu\text{m}$
40	1.3 (oil)	0.12 $\mu\text{m}$
63	1.4 (oil)	0.11 $\mu\text{m}$
100	1.4 (oil)	0.11 $\mu\text{m}$



Values are for scan zoom = 1.0

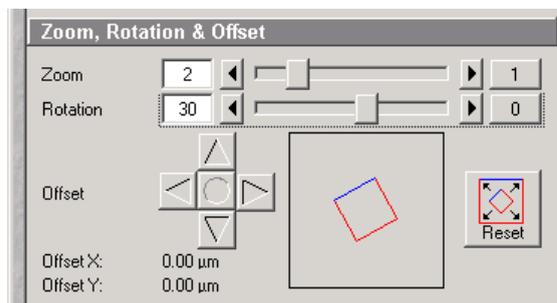
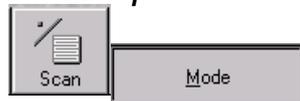
Adjusting the field size ("XY") to 56  $\mu\text{m}$  with the 63x lens, would produce a pixel size of 0.1  $\mu\text{m}$

Brightness of image =  $\text{Magnification}^2 / \text{NA}^2$

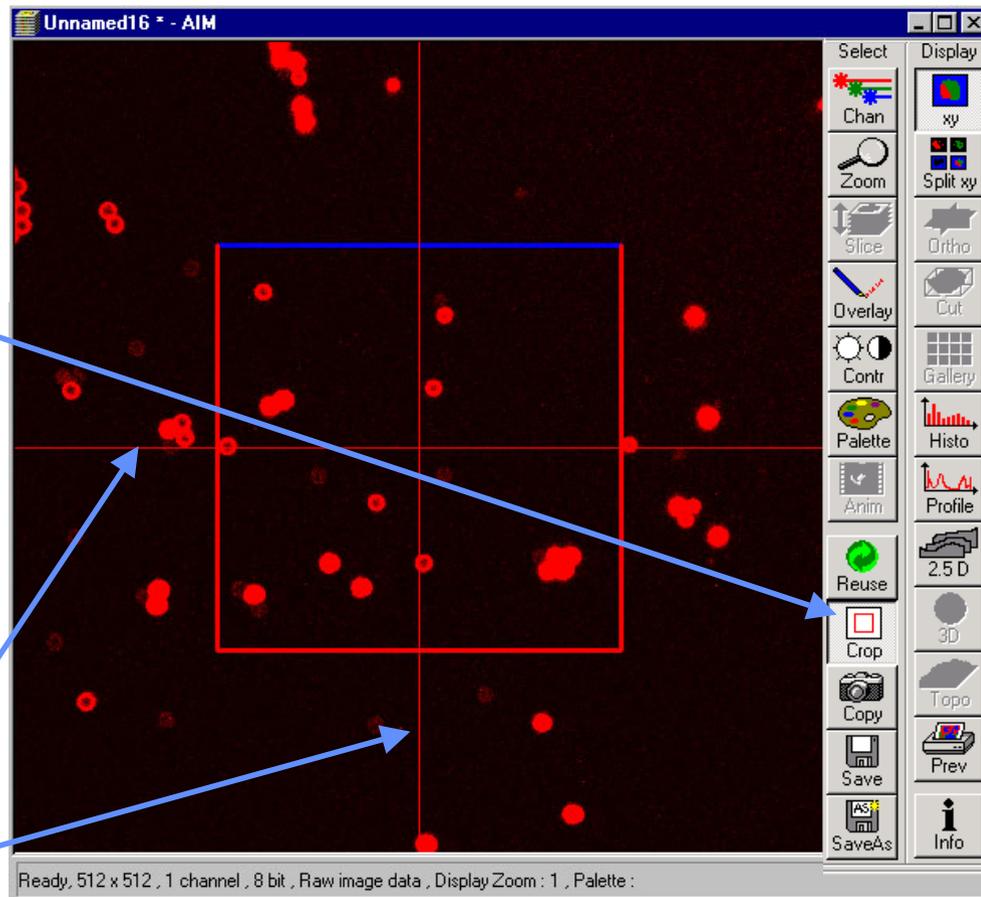
Field size can be adjusted by changing the objective magnification, or by optical zooming. Changing from 63  $\times$  to 100  $\times$  will reduce the field size, but will also reduce the amount of light available.

## Optical Zooming

The level of zoom can be changed either by using the *Zoom, Rotation & Offset* control in *Mode* menu of the *Scan Control*, or by selecting *Crop* in the image menu.



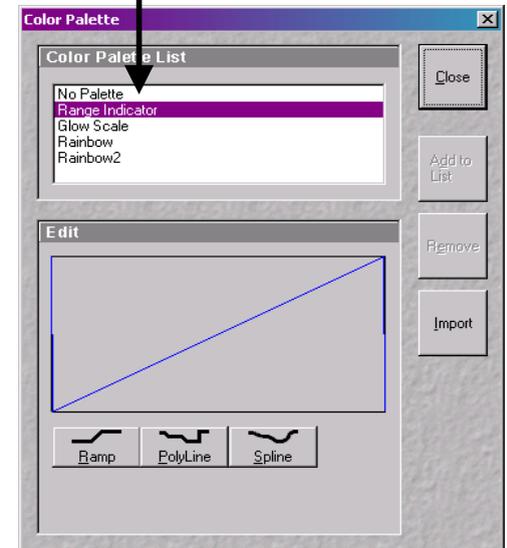
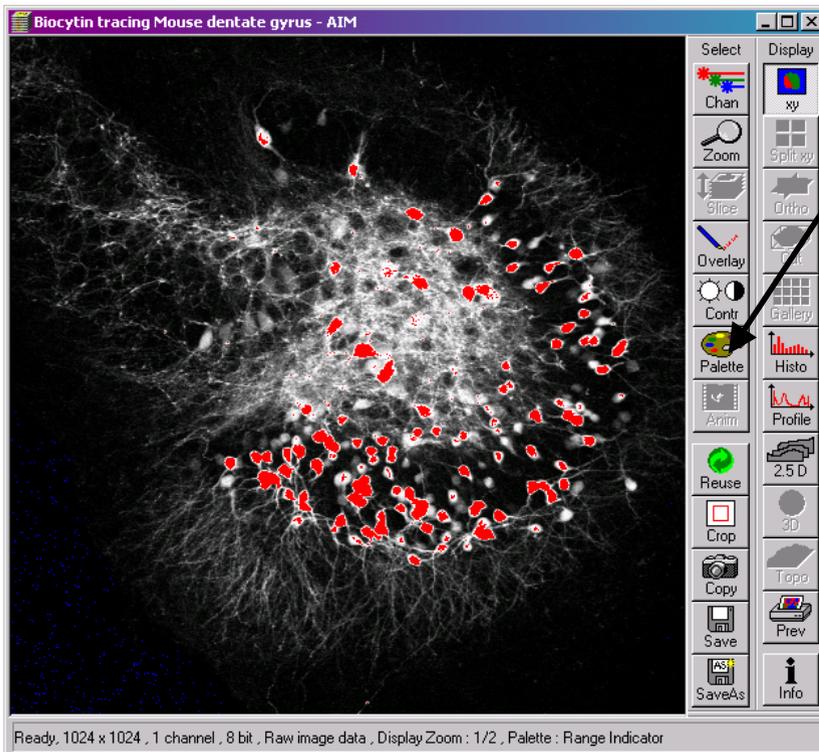
The image can also be rotated by selecting and dragging the bars



## Selecting gain and offset – Choosing a lookup table

1) Select *Palette*

2) Select Range Indicator



Red = Saturation (maximum)

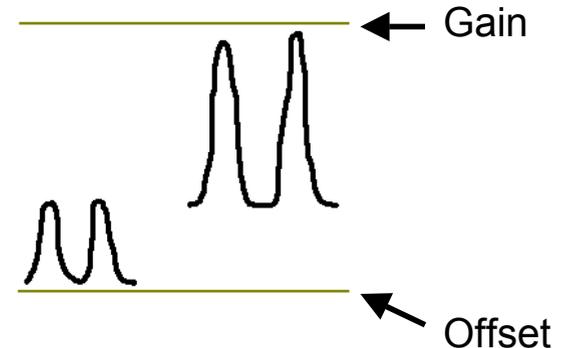
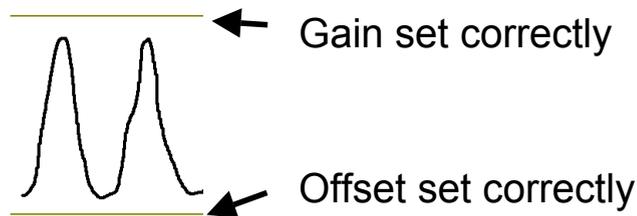
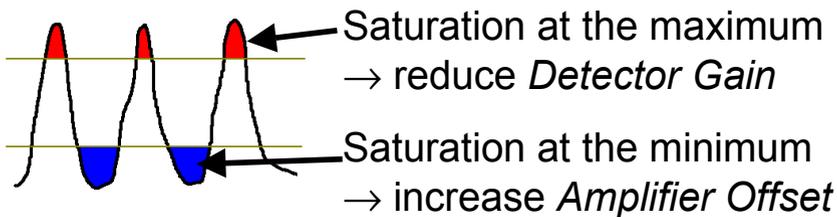
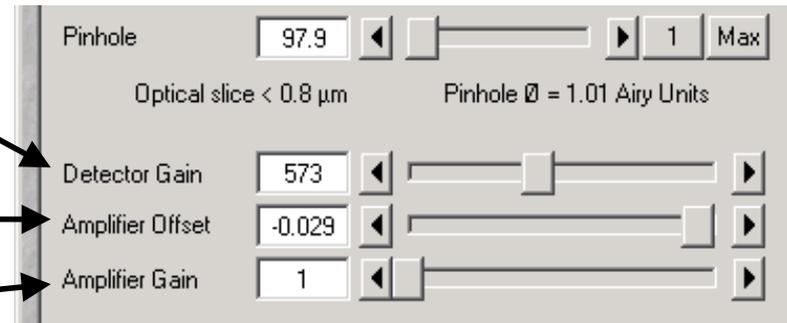
Blue = Zero (minimum)

## Scan Control – Setting Gain and Offset

*Detector gain* determines the sensitivity of the detector by setting the maximum limit

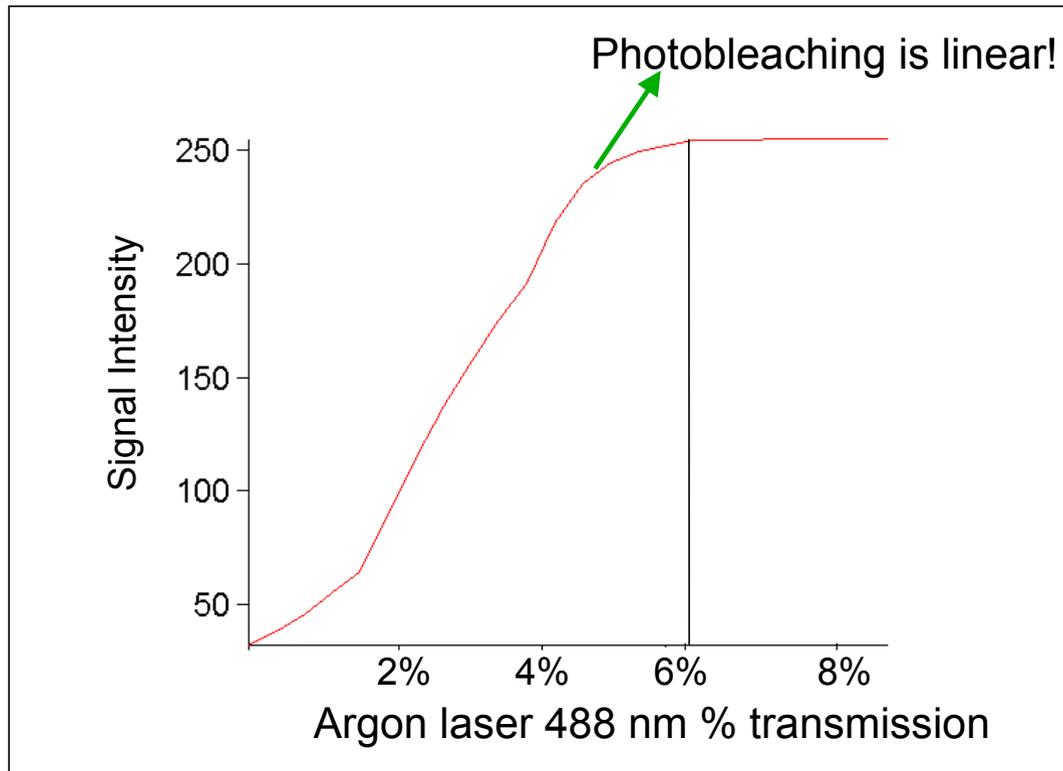
*Amplifier Offset* determines the minimum intensity limit

*Amplifier Gain* determines signal amplification



*Amplifier Gain* increases the whole signal, and the *Amplifier Offset* will need to be decreased.

## Saturation of Signal Intensity with Laser Power



- Fluorophore saturates at 6% laser transmission
- Photobleaching is linear

Laser transmission should not be set higher than the saturation level.

## Adjusting the Laser Intensity

- 1) Set *Pinhole* to 1 Airy unit
- 2) Set *Detector Gain* high
- 3) When the image is saturated, reduce AOTF transmission in the *Excitation* panel to reduce the intensity of the laser light at the specimen

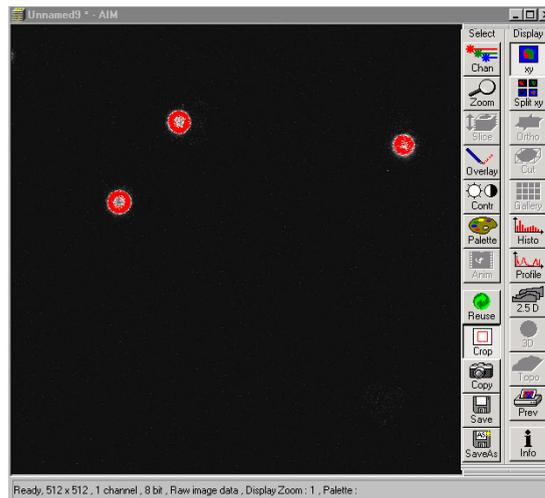
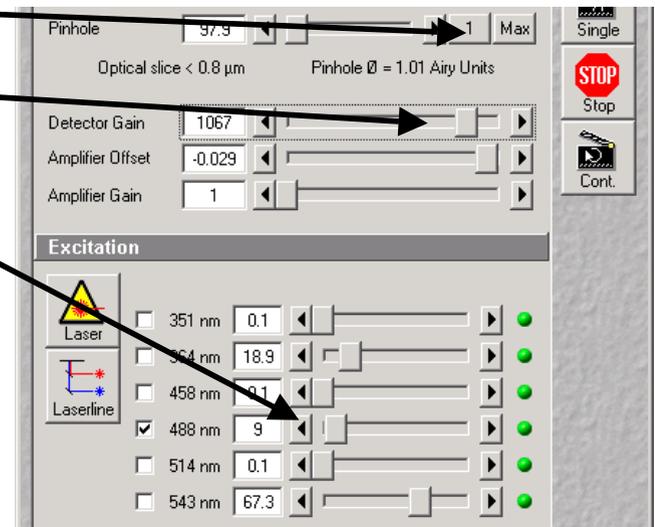
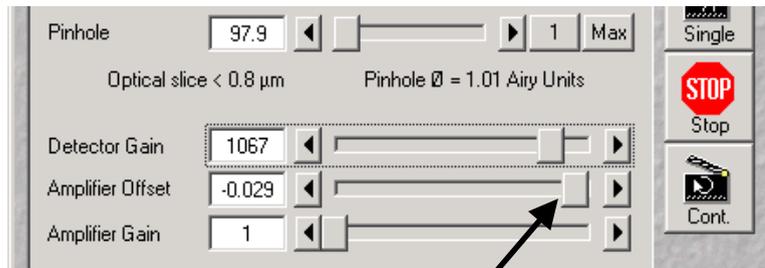
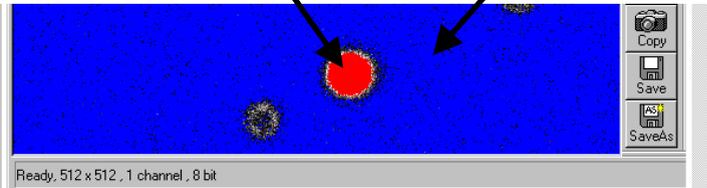


image with saturated pixels

## Adjusting Gain and Offset



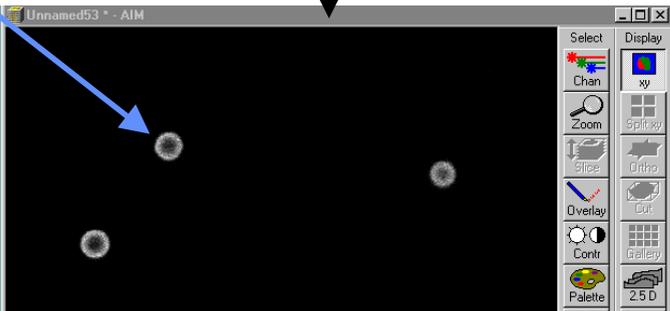
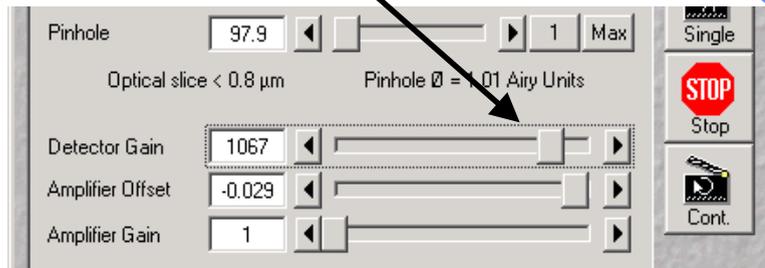
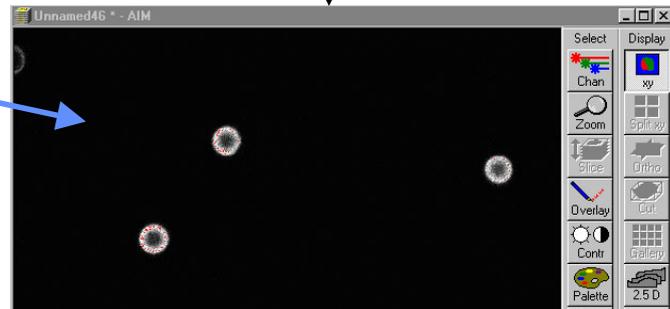
Both *Detector Gain* and *Amplifier Offset* saturated



gain and offset **not** correct

1) Increase the *Amplifier Offset* until all blue pixels disappear, and then make it slightly positive.

2) Reduce the *Detector Gain* until the red pixels only just disappear.



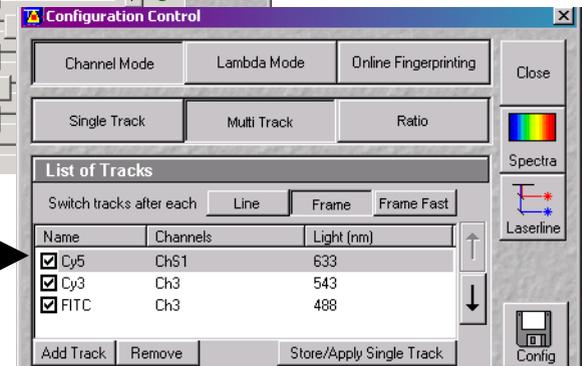
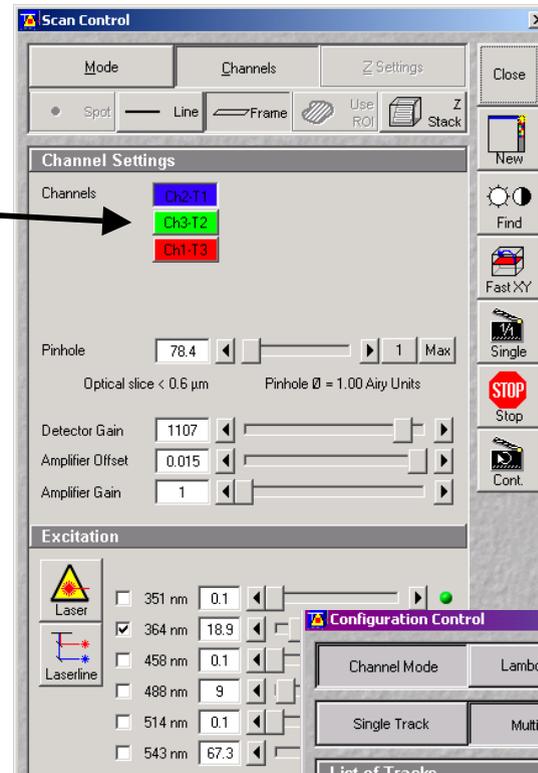
gain and offset correct

no blue  
no red

## Adjusting the Laser, Gain and Offset using a Multi Track Configuration

Each channel is selected independently by clicking on the colour button indicating the channel i.e. *Ch2-T1* (Channel 2, Track 1). The laser power and all other parameters are optimised as described in the previous slides for each selected channel.

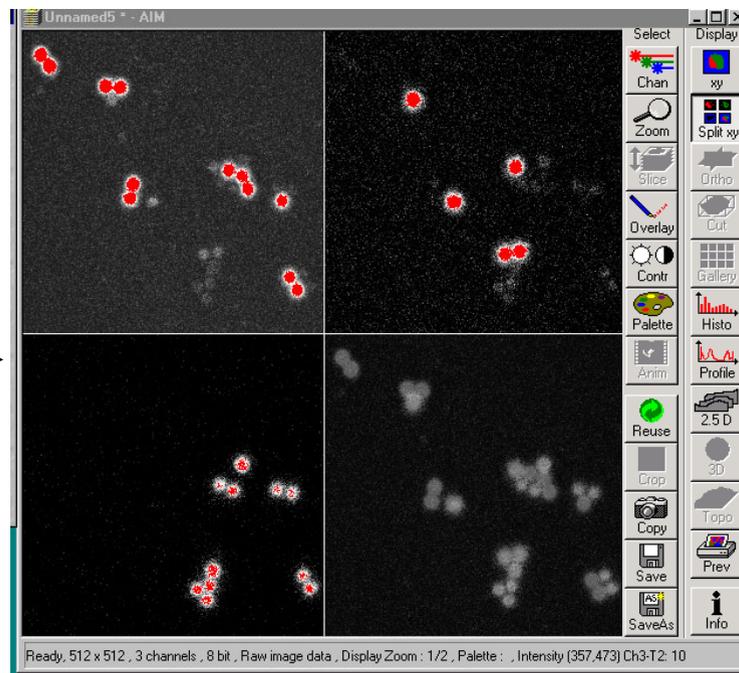
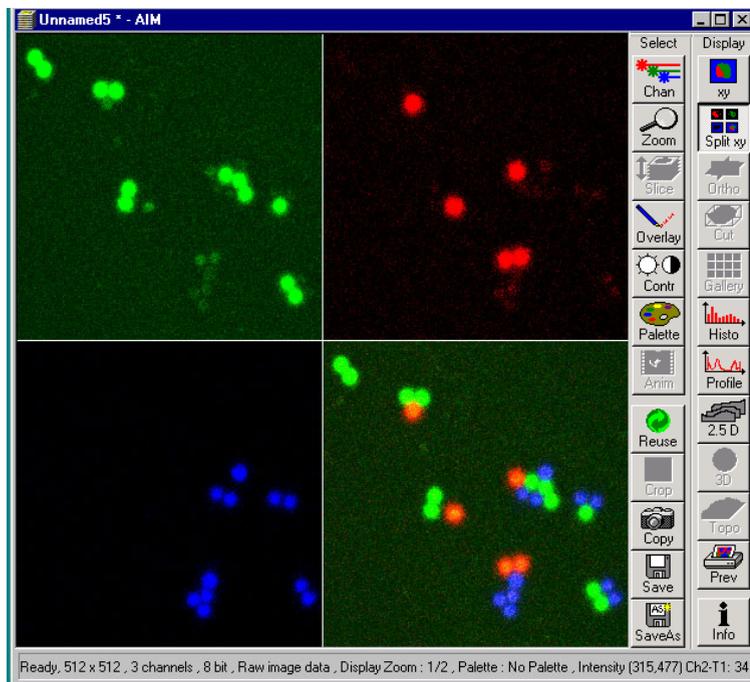
For accurate colocalisation, adjust each *Pinhole* so that each channel has the same *Optical Slice*



To adjust laser, gain or offset for a single track in a multi-track configuration it is possible to temporarily deactivate the other tracks in the *Configuration control*

## Setting up Gain and Offset - Multi Track

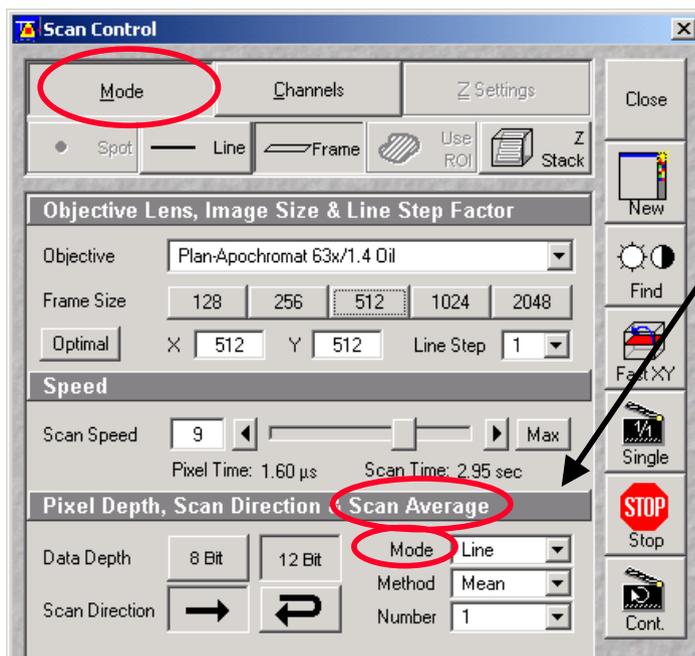
- 1) Select *Split XY* in the Image window
- 2) In *Palette*, select *Range indicator*
- 3) Select each channel separately under *Channels* in the *Scan control* window and adjust the *Laser intensity*, *Detector Gain*, and *Amplifier Offset* as described previously.



## Line Averaging

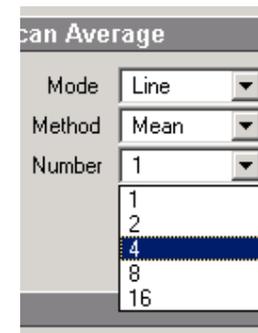
Averaging improves the image by increasing the signal : noise ratio

Averaging can be achieved line by line, or frame by frame



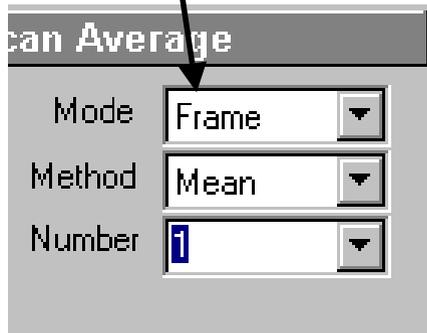
1) Select *Line* or *Frame* under *Mode* in *Scan Average* within the *Mode* panel of the *Scan Control* window

2) Select *Number* for averaging. The more the better for the signal to noise ratio (max 16) in this case, each line will be scanned 4 times. But: Averaging increases the exposure time of the sample!!

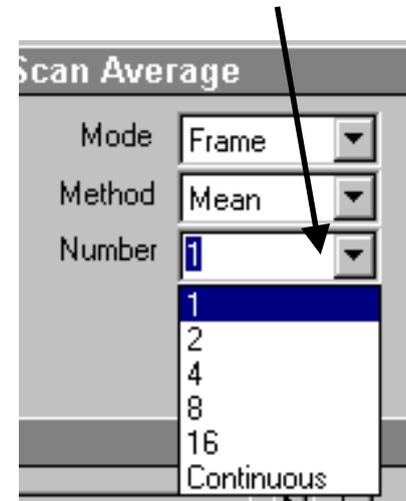


## Frame Averaging

1) Select *Frame*

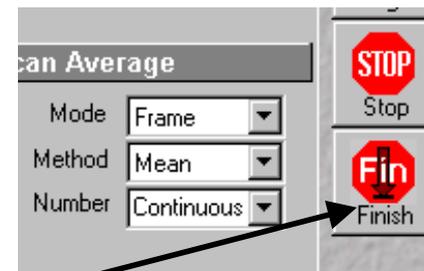


2) Select the *Number* for averaging - The more the better for signal to noise ratio (max 16). Continuous averaging is possible in this mode.



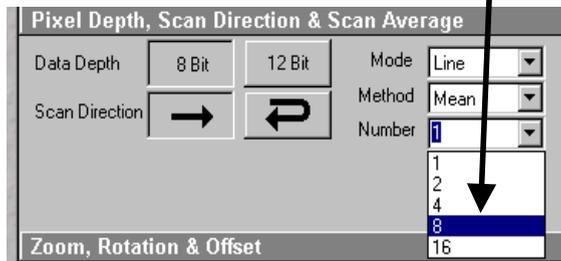
Frame averaging helps reduce photobleaching, but does not give quite such a smooth image. There is also a longer delay between each track when using "Multi Track".

Continuous averaging has a *Finish* button which allows the scan currently in progress to be completed before stopping

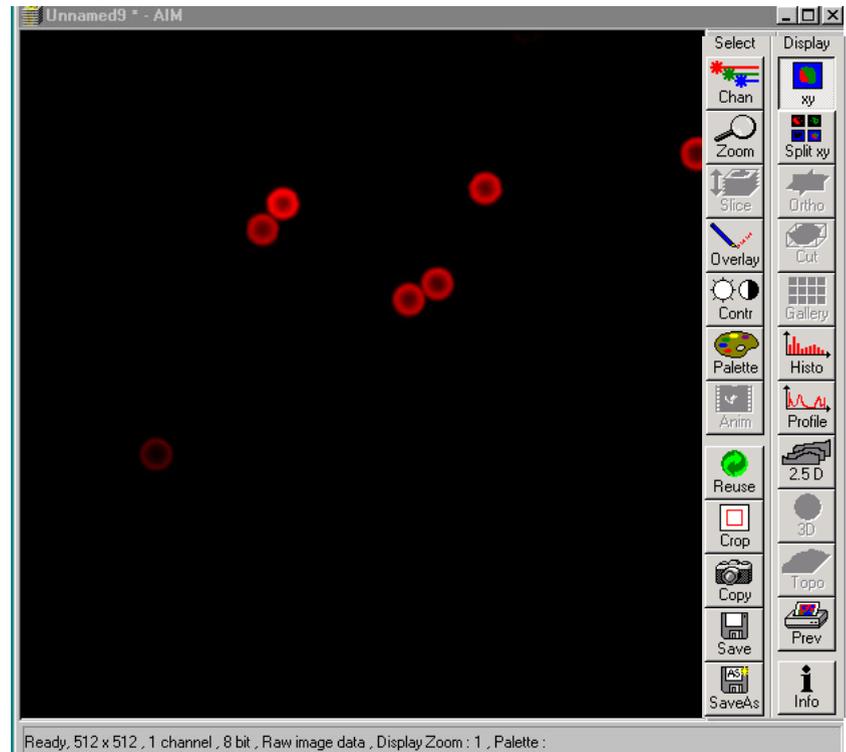
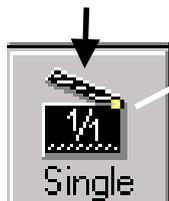


## Collecting an Averaged Image

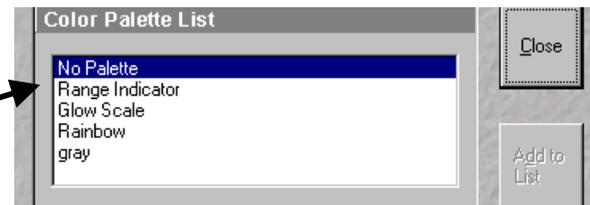
1) Under *Scan Average* select the *Number* for the average.



In the *Channels* panel of the *Scan Control* window select *Single*. An averaged image will be collected.



Range indicator set to *No Palette*



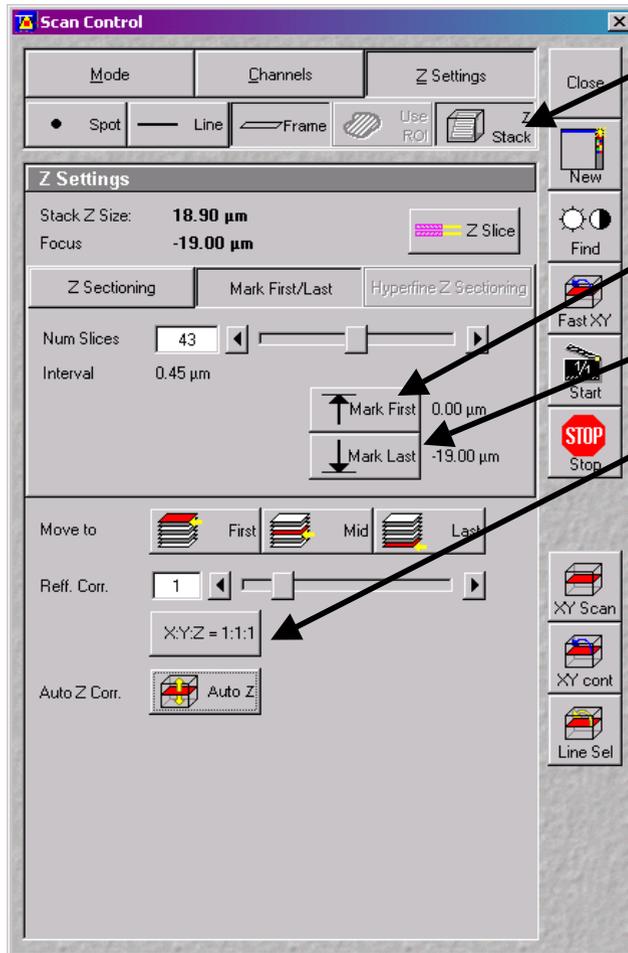


## Contents

- Starting the Zeiss LSM 510 microscope, software and laser  
Selecting an objective and focusing the microscope
- Selecting an objective and focusing the microscope
- Configuring the laser scanning and detection for confocal image acquisition
- **Acquiring a Z- and Time - Series**
- Data storage

Descriptions also include the LSM 510 META

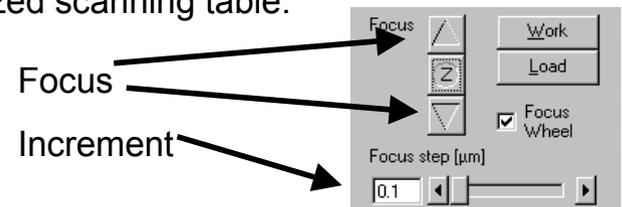
## Scanning a Z-Series using *Mark First/Last*



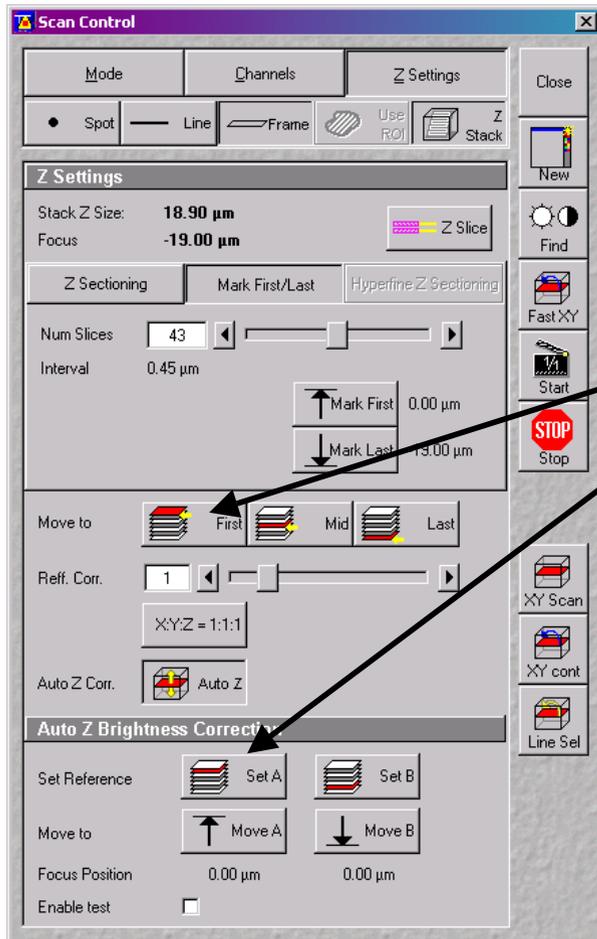
- 1) Select *Z Stack*
- 2) Start scanning using *Fast XY* or *XY cont*
- 3) Keep your eye on the image and move the focus to the beginning of the Z-Series, then select *Mark First*
- 4) Move the focus back in the opposite direction to the end of the Z-Series, then select *Mark Last*
- 5)  $X:Y:Z = 1:1:1$  sets the Z-interval so that the voxel has identical dimensions in X, Y, and Z.
- 6) *Start* will initiate the acquisition of the Z-Stack. The acquisition can be stopped at any time.

### NOTE

Focusing can be achieved manually (preferred), or using *Stage* in the LSM menu if there is a motorized scanning table.



## Using Auto Z Brightness Correction



*Auto Z* provides an automatic gradual adjustment of *Detector Gain*, *Amplifier Offset*, *Amplifier Gain*, and *Laser intensity* setting between the first and last optical slice of a *Z Stack*.

1) After defining the *Z* position of the first and last optical slice activate *Auto Z*.

2) Move to the *First Slice* and adjust the parameter for the image acquisition in the *Channels* panel for each used channel as described in the previous slides. Then click on *Set A* to store the values.

3) Repeat the procedure after moving to the *Last Slice*. Click on *Set B* to store the parameters for the last slice.

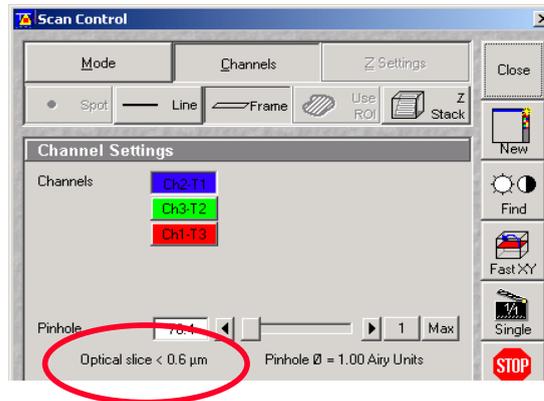
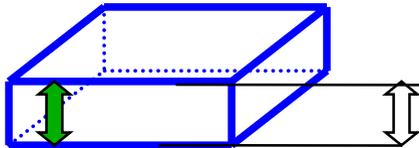
**Note:** Positions *A* and *B* do not have to be the first and last slice of a stack and can also be defined simply by focussing to the appropriate positions, adjusting the parameters and pressing *Set A* or *Set B*.

4) The parameters for image acquisition will be gradually and linearly adjusted between the first and last slice of the *Z Stack*. Thus signal intensity and image quality is comparable throughout the *Z Stack*.

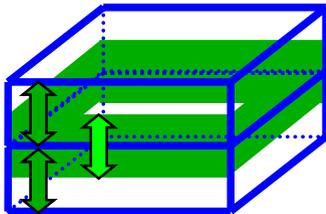
## Confocal Z Sectioning Number of Sections for correct sampling

Optical thickness  $d$  depends on:

- Wavelength  $\lambda$
- Objective lens,  $N.A.$
- Refractive index  $n$
- Pinhole diameter  $P$



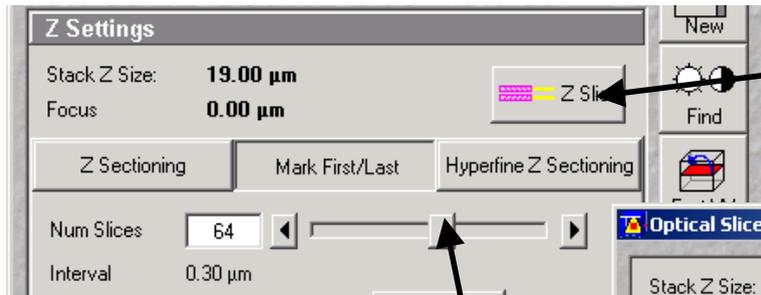
The optical slice thickness is displayed in the *Scan Control*



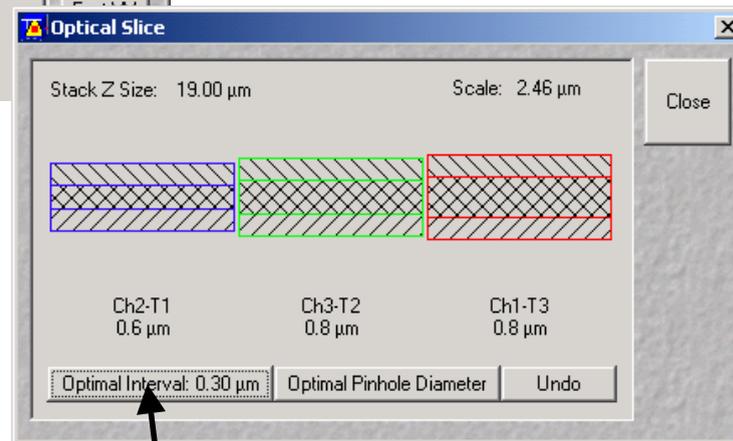
For Z-sectioning it is optimal to have:  
no missing information @ minimal number of sections  
*Slices overlap by the half of their thickness*

**„Nyquist-“ or Sampling-Theorem**

## Z Stack – Number of Slices and Increment



1) Select *Z slice* - the window *Optical Slice* will appear



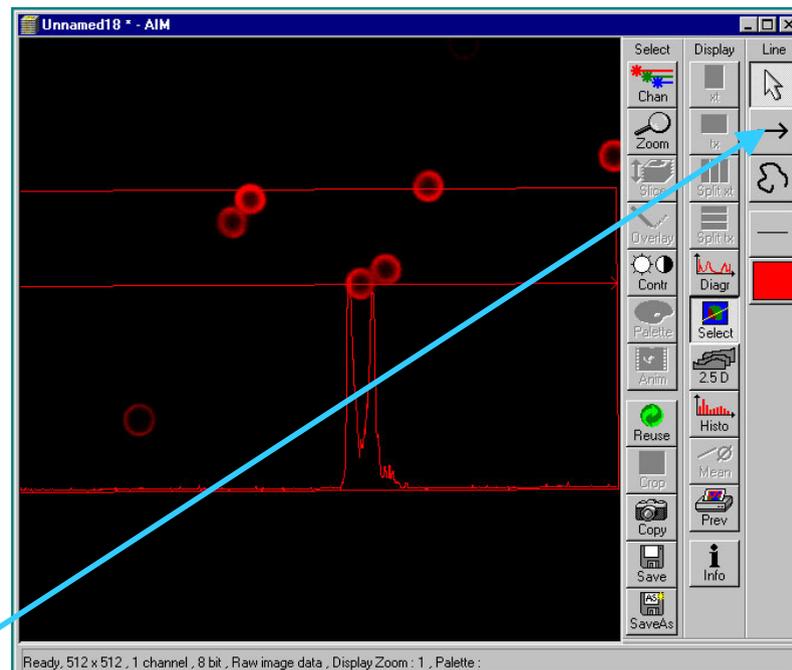
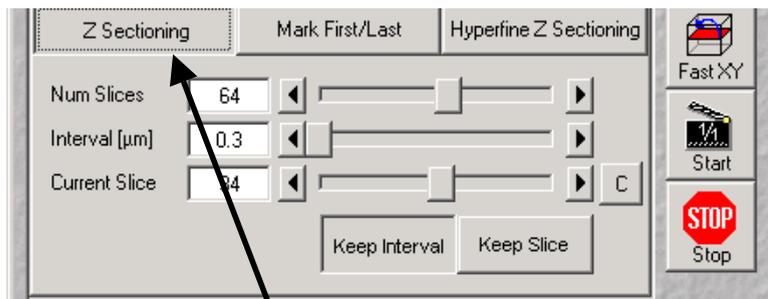
2) Select *Optimal interval* the computer will calculate the optimum number of sections

For more or less sections - adjust *Num Slices*

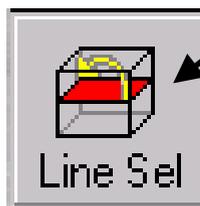


3) Select *Start* to acquire the complete stack

## Z - Series using Z Sectioning

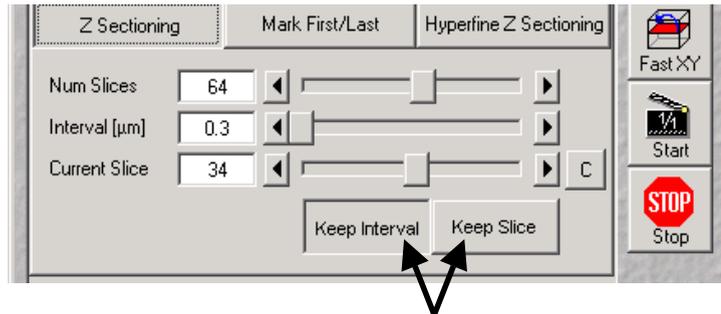


- 1) Select *Z Stack*
- 2) Select *Z Sectioning*
- 3) Select *Line Sel*
- 4) Select the large arrow button and position the XZ cut line



XZ outline will be displayed as diagram within the XY image

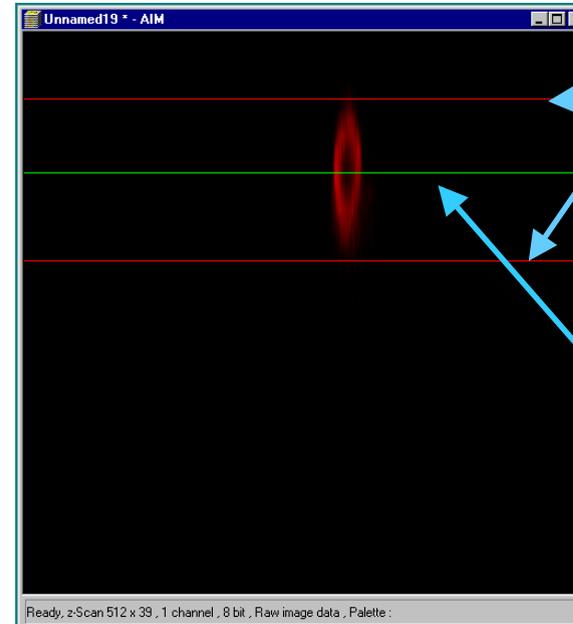
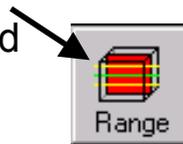
## Z Sectioning – Setting Range



1) Decide whether to *Keep Interval* (number of slices will change) or *Keep Slice* (Interval between slices will be adjusted)

2) Select *Range* and position bars to decide where the Z - Series begins and ends

3) Select *Start* for image acquisition

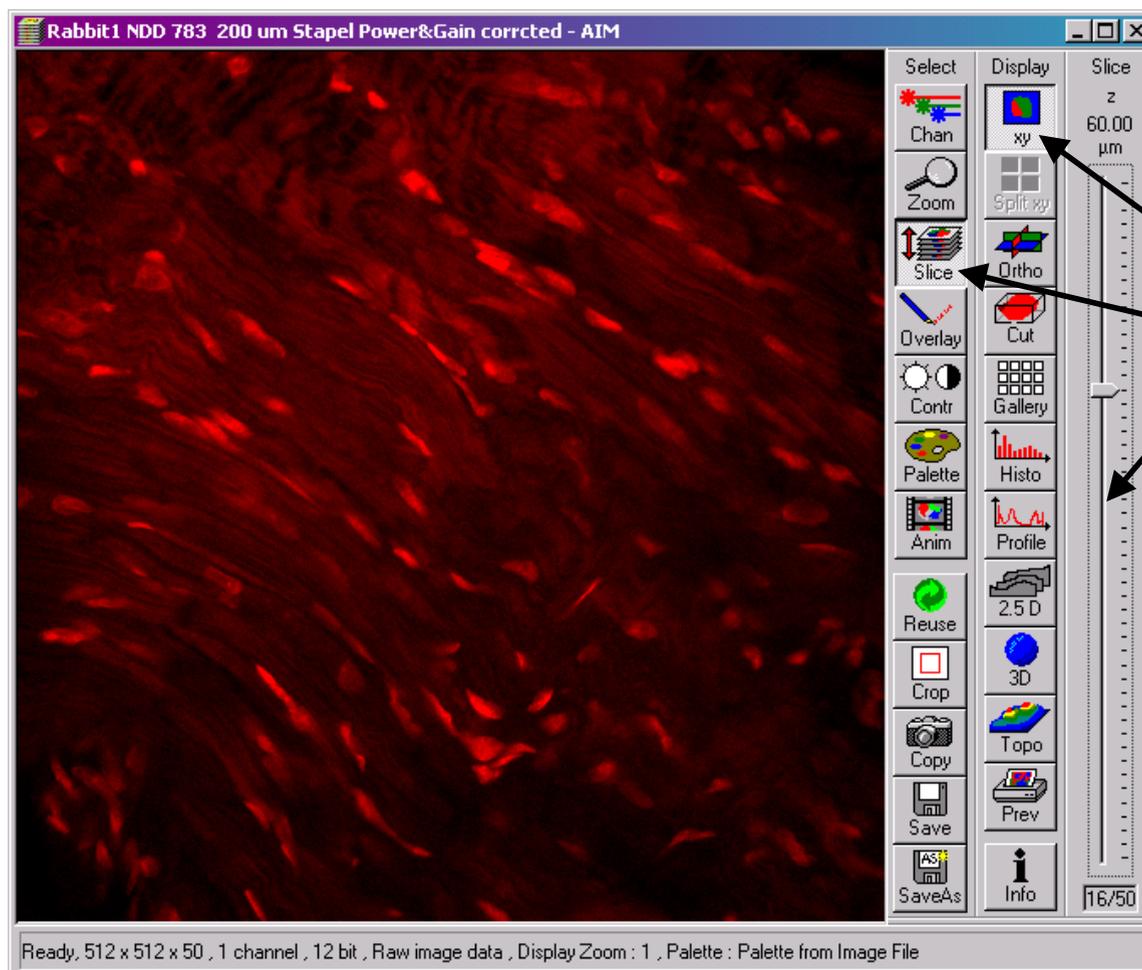


Pull red lines to set limits for Z-Series

Drag green line to change focus position

Pressing *Range* produces an XZ image of selected Z-range, plus 50% above and below the selected stack.

## Viewing a Z - Series



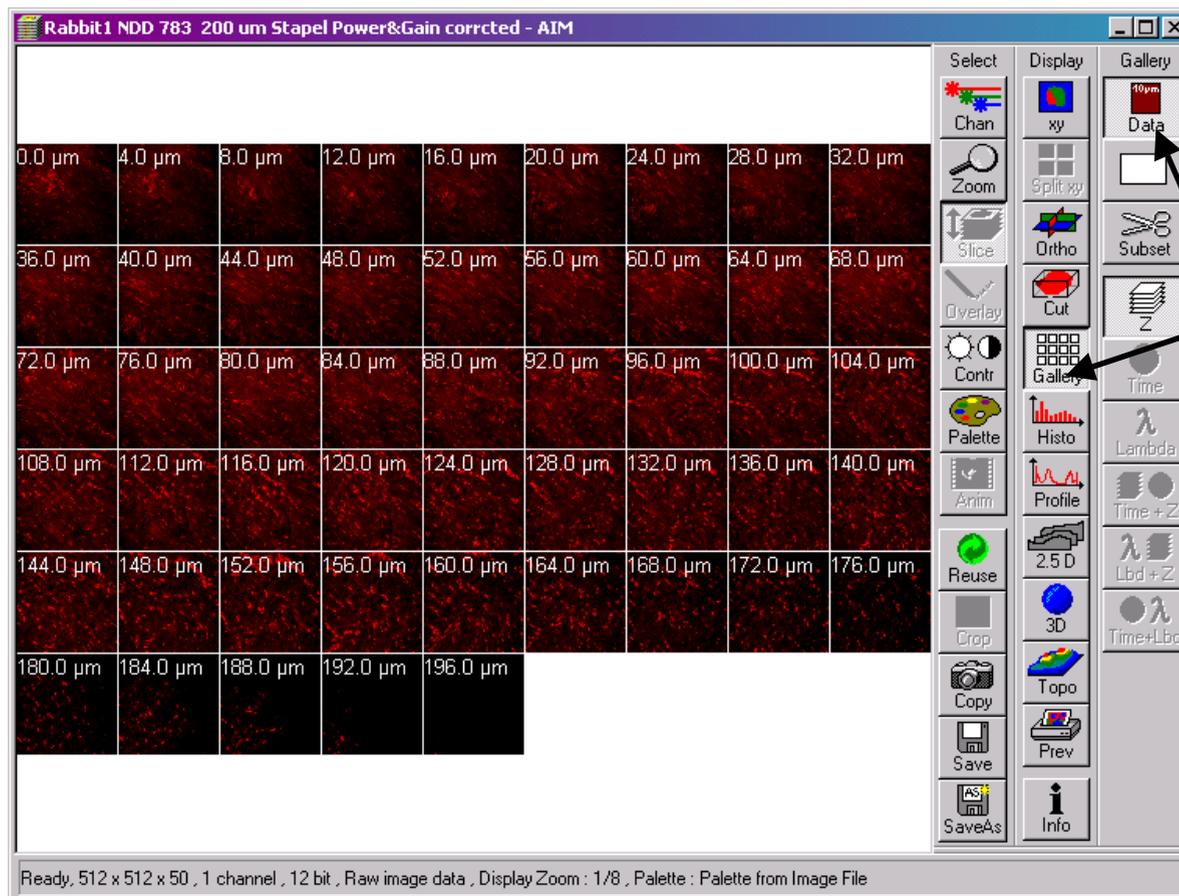
In the image window

1) Select xy

2) Select *Slice*

3) Use scroll bar to view individual sections

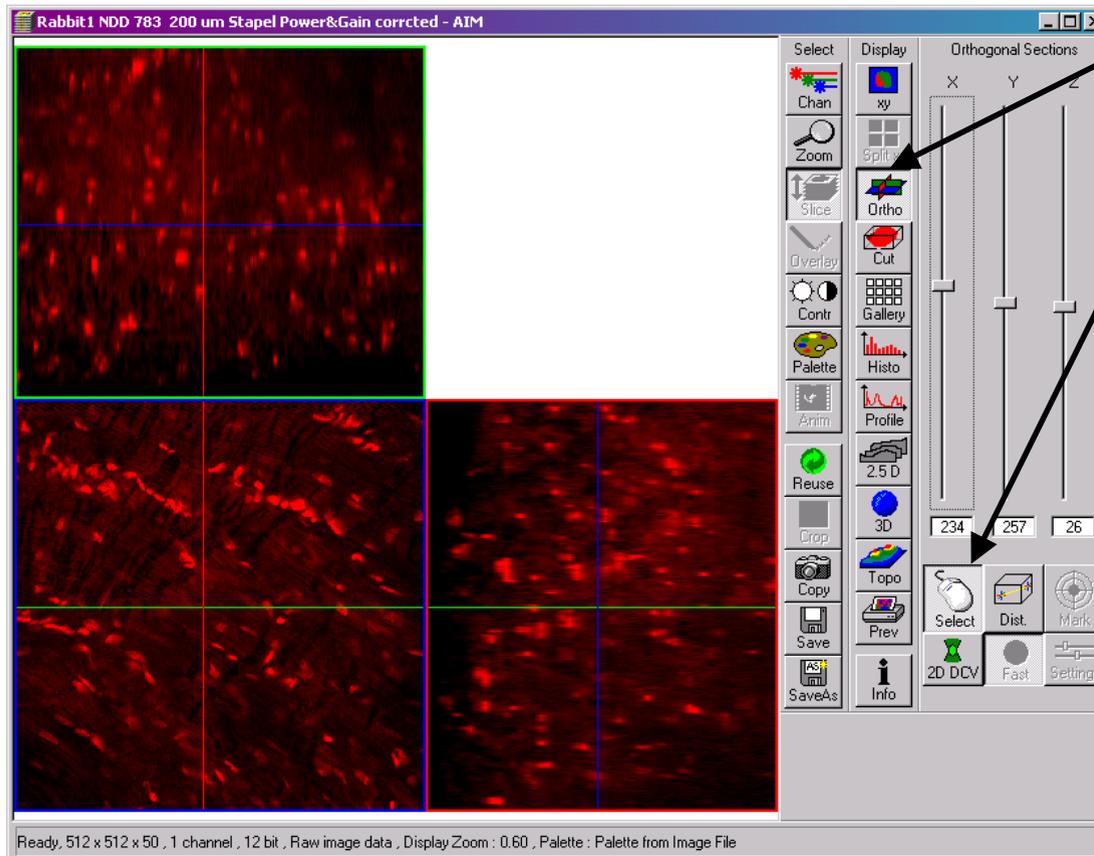
## Viewing a Z - Series using *Gallery*



- 1) Select *Gallery*
- 2) Select *Data* for scale

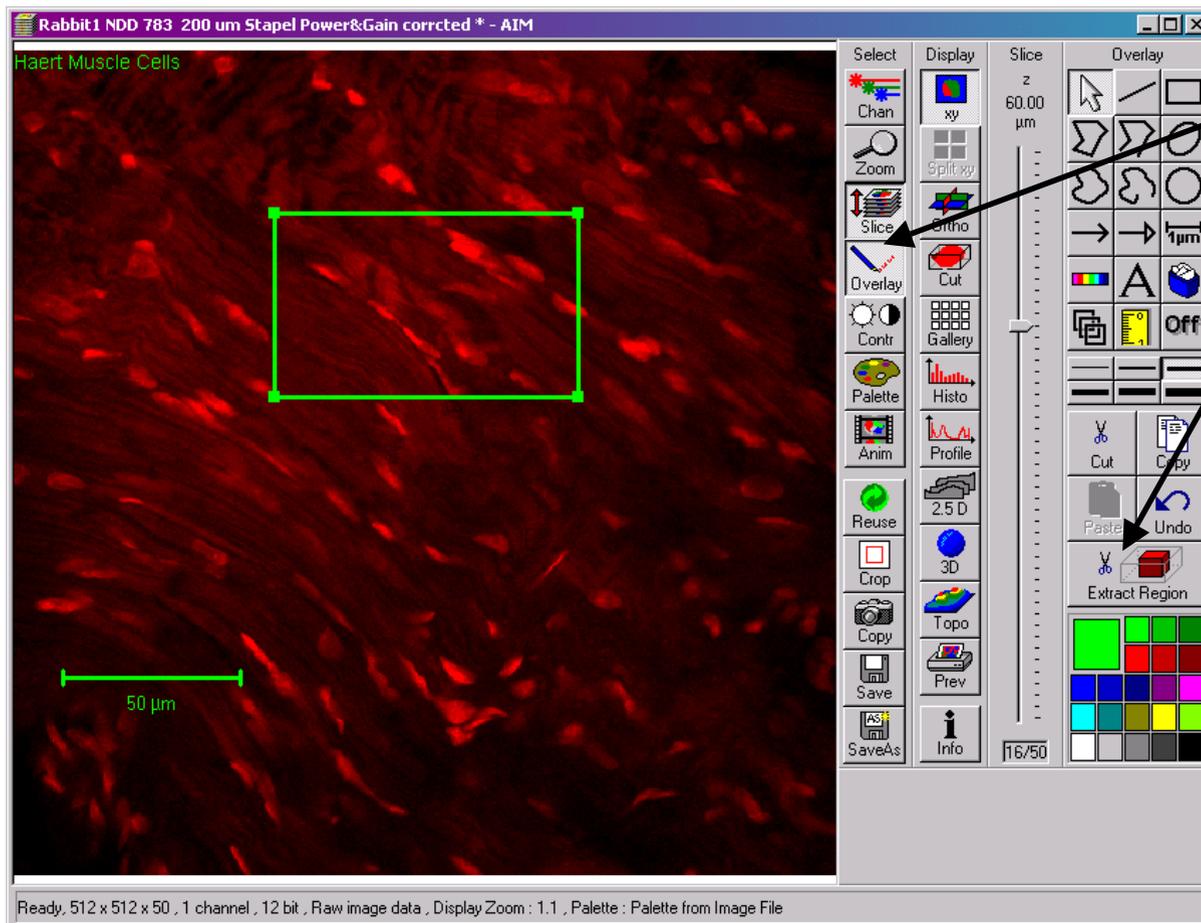
Use *Subset* to extract sections

## Viewing a Z- Series using Orthogonal Sections



- 1) Select *Ortho*
  - 2) Select mouse (*Select*)
  - 3) Using the mouse, position the cut lines.
- To save orthogonal sections, select *Export* and save as *contents of image window*.

## Selecting and Saving a Region of Interest (ROI)

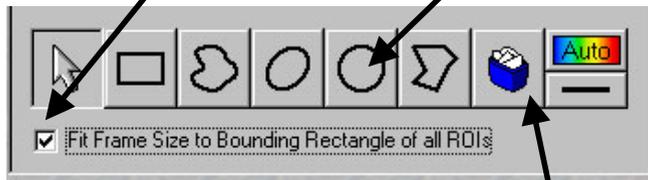


- 1) Select Overlay and define shape of ROI
- 2) Extract region creates a Z-Stack from the ROI
- 3) Save data

## Using a ROI for faster image acquisition and data saving



- 1) Select *EditROI* from the LSM menu bar
- 2) Select *Fit Frame Size to bounding Rectangle*
- 3) Choose shape of ROI

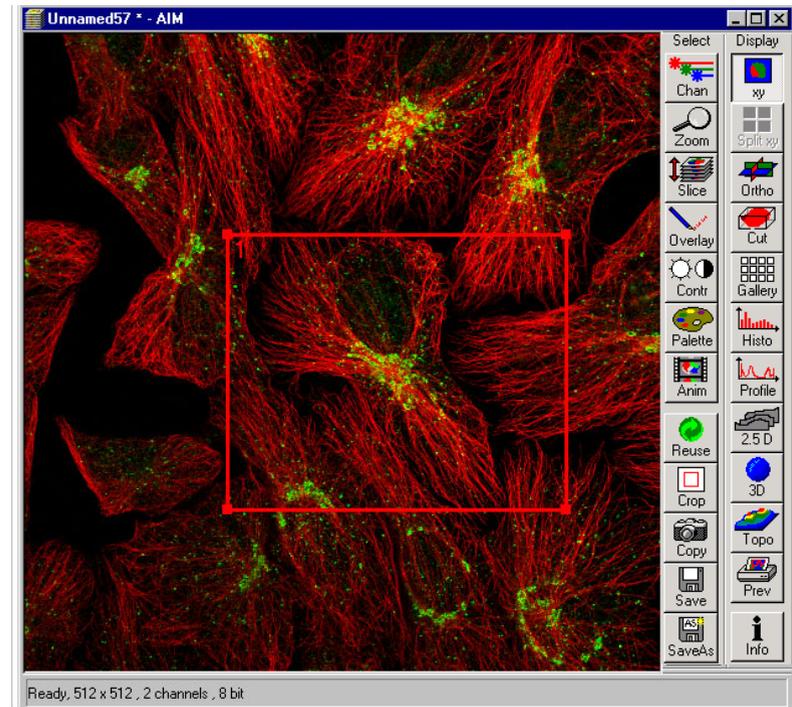


4) Position and size the ROI in the image with the mouse

5) Start Scan

Interactive ROI Definition					
	Type	Center Position		Dimension	
		X	Y	X	Y
<input type="checkbox"/> 1	Ellipse	210	234	104	126
<input checked="" type="checkbox"/> 2	Bezier	249	311	224	189

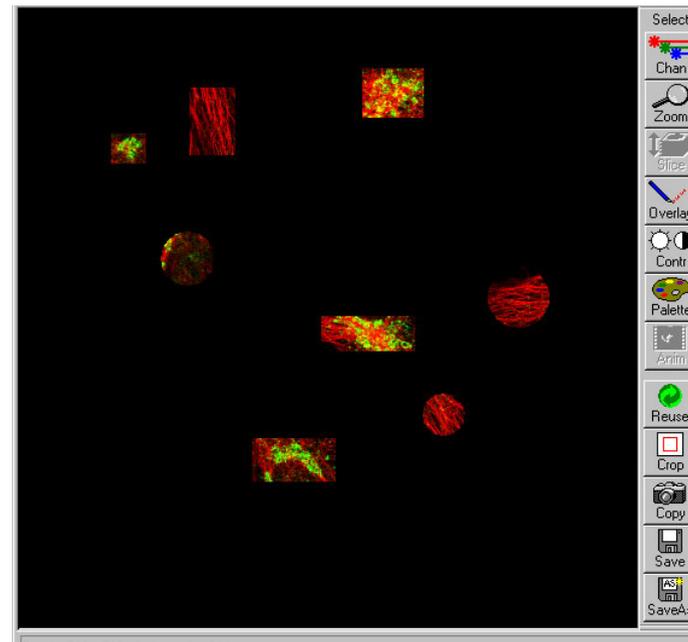
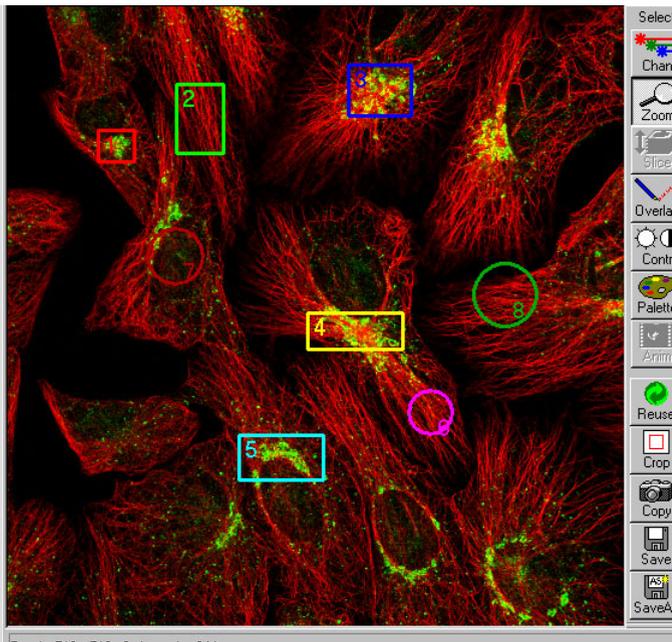
To remove ROI and overlay select blue bin or deactivate ROI. Closing the window only removes overlay, ROI is still active. Deactivate *Use ROI* in the LSM menu.



## Multiple Regions of Interest

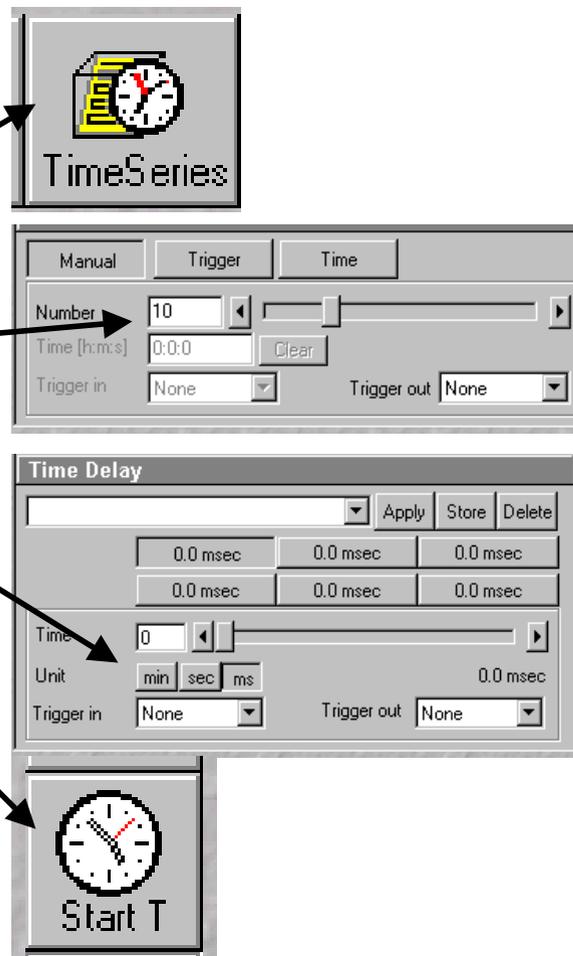
- 1) Un-select *Fit Frame Size to bounding Rectangle*, Choose shapes of ROIs
- 4) Position and size the ROIs with mouse
- 5) Start Scan

To remove ROIs and overlay select blue bin or deactivate ROIs. Closing the window only removes overlay, ROIs are still active. Deactivate *Use ROI* in the LSM menu.



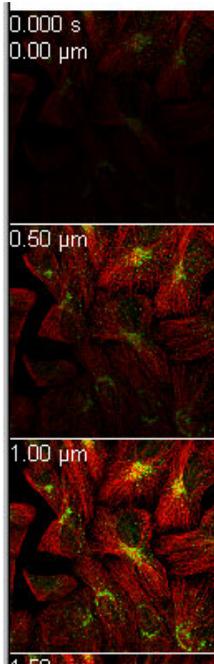
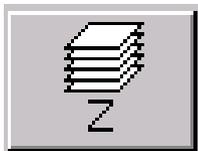
## Time Series

- Set up scanning parameters for image acquisition as described in previous slides
- Select *TimeSeries* from the LSM menu
- Enter the *Number* of cycles
- For a Time Delay between image acquisition select *min*, *sec* or *ms* and set time with the slider
- Select *Start T* to start image acquisition
- Instead of using *Manual* you can select *Time* to start and stop the series at a certain system time!

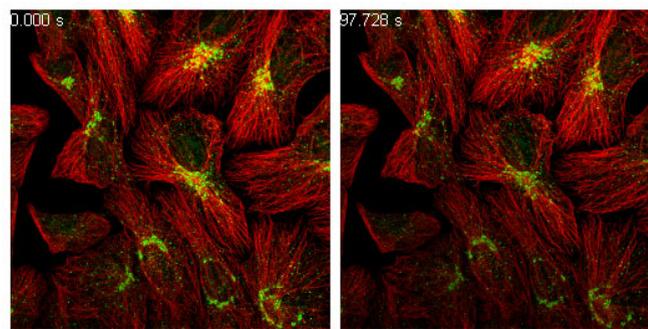
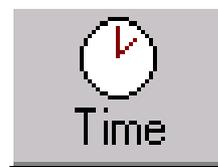


## Viewing a Time Series of a Z Stack

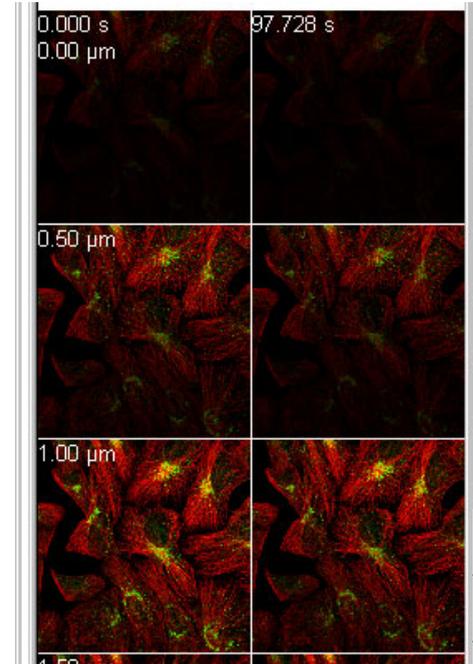
Z Sections  
for any time



Time points for  
any Z Section

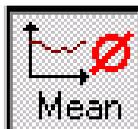


Both Z sections  
and time series



## Time Series – Physiology Experiments

- 1) If required, use multiple regions of interest
- 2) Set up Time Series as before
- 3) Instead of using ~~StartT~~ select MeanROI to start scanning



View and save data by selecting *Mean* in the image window

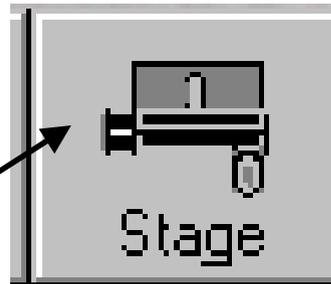
The screenshot displays five intensity time series plots (Intensity HUI 1, Intensity ROI 2, Intensity ROI 3, Intensity ROI 4, Intensity ROI 5) and a control panel on the right. The control panel includes a 'Scan Mean of ROIs' table, a 'Diagram' section, and a 'Scaling' section.

Scan Mean of ROIs						
Type	Position X	Position Y	Dimension X	Dimension Y		
<input checked="" type="checkbox"/>	1	352	132	106	124	
<input checked="" type="checkbox"/>	2	252	100	125	103	
<input checked="" type="checkbox"/>	3	223	307	114	79	
<input checked="" type="checkbox"/>	4	359	309	110	154	
<input checked="" type="checkbox"/>	5	95	398	172	174	

The control panel also includes a 'Diagram' section with buttons for '1', 'Chan', 'ROI', and 'Mono'. The 'Scaling' section includes buttons for 'Automatic', 'Time Range', and 'Number Cycles'. The 'Image / Table' section includes buttons for 'Show Image', 'Copy Table', 'Show Table', and 'Save Table'.

## Imaging a large area using *Tile Scan*

This function is only available with a motorized stage



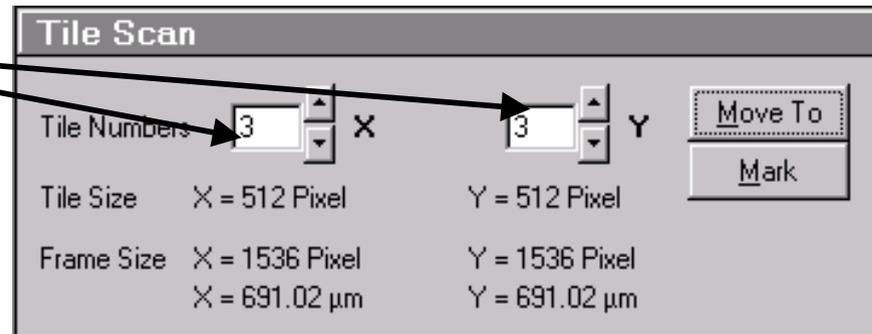
1) Select *Stage* on LSM menu

2) Enter the *Tile Numbers*

3) Select *Start*

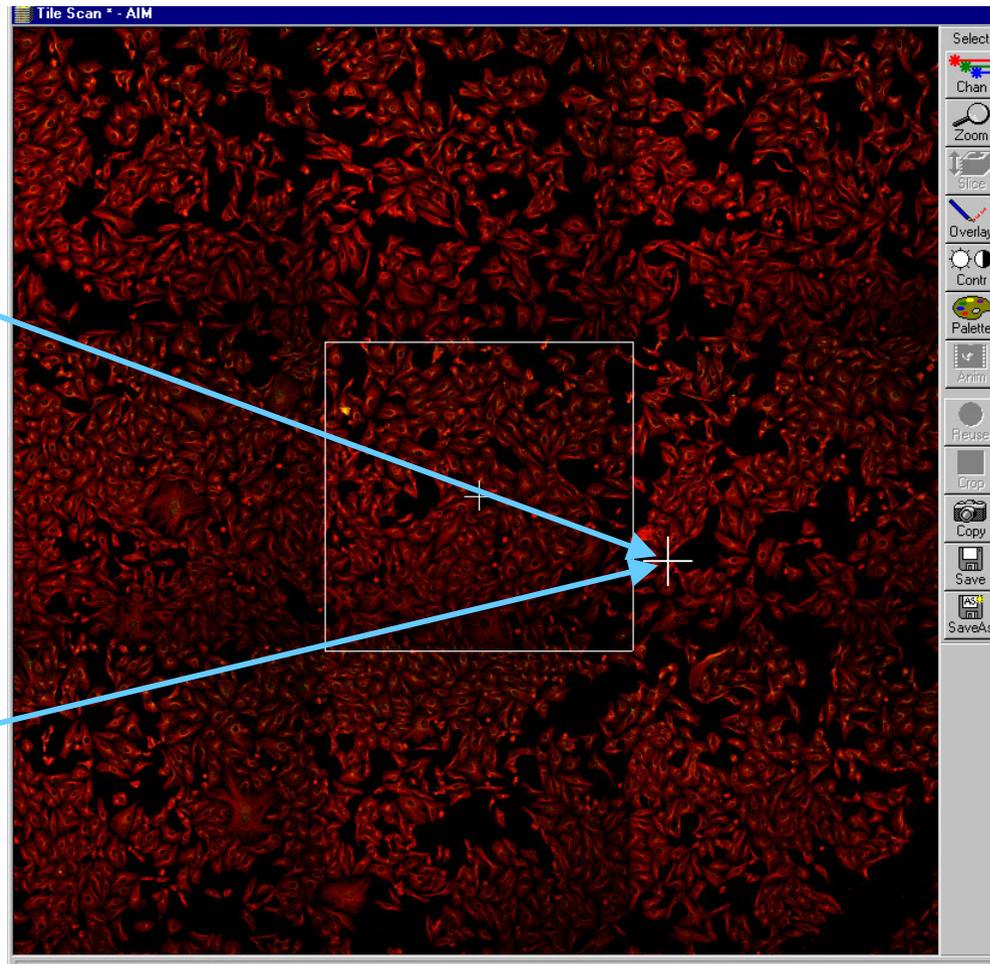
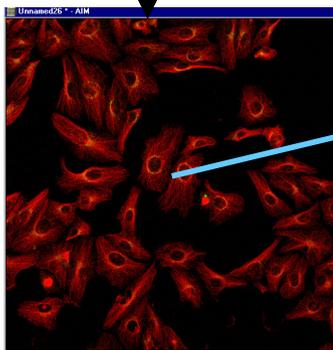
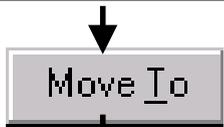


The maximum size is 4096 x4096 pixels



## Tiled Image

Any position can then be marked and a single image acquired by selecting *Move to* and then single





## Contents

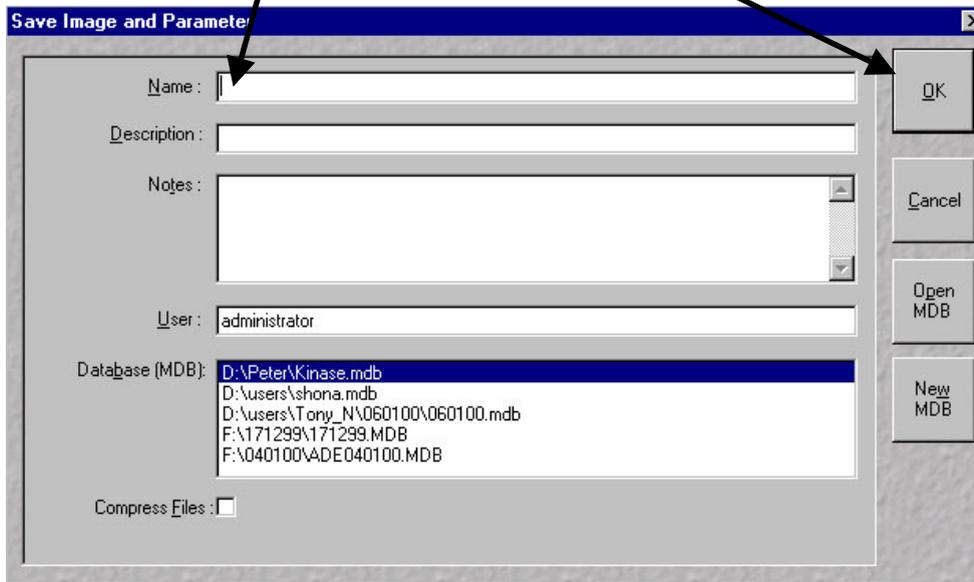
- Starting the Zeiss LSM 510 microscope, software and laser  
Selecting an objective and focusing the microscope
- Selecting an objective and focusing the microscope
- Configuring the laser scanning and detection for confocal image acquisition
- Acquiring a Z- and Time - Series
- **Data storage**

Descriptions also include the LSM 510 META

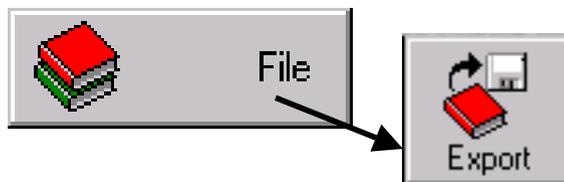
## Saving Data - Using Database



- 1) Select Save or Save as on image window or LSM menu bar
- 2) Enter file name and notes if required
- 3) Select OK

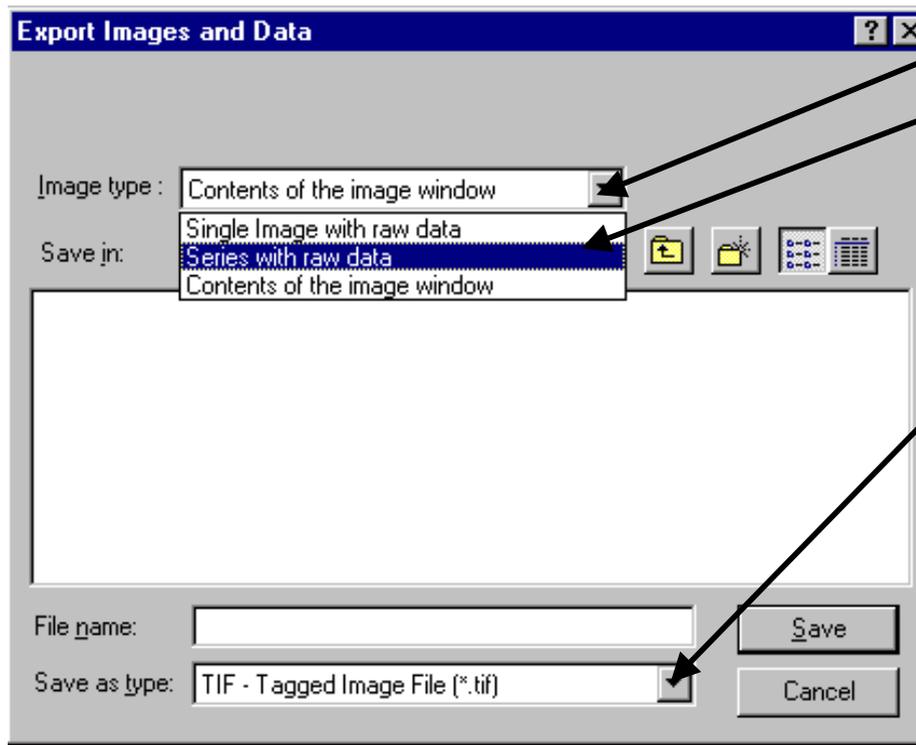


## Saving Data – Using *Export*



1) Select *File* from LSM menu

2) Select *Export*



3) Select *Image type*

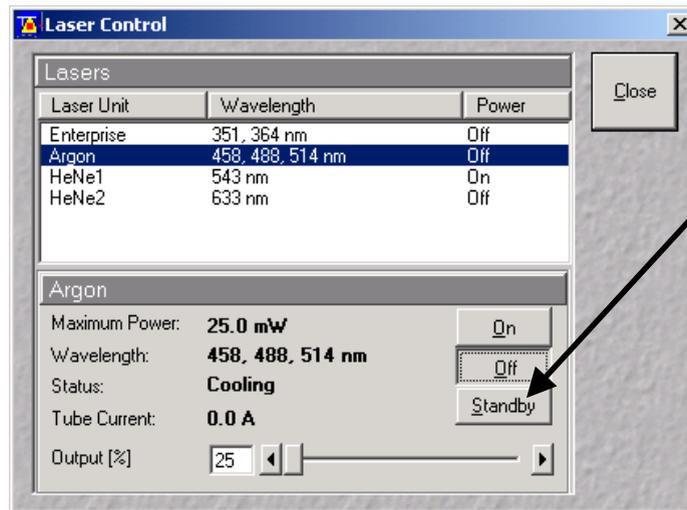
4) Select *Single image with raw data* (No overlay or Look up table etc. is saved), *Series with raw data*, or *Contents of the image window* (Saves the image as shown on the screen)

5) Select *Save as type*

Tif - Tagged image File” is OK for 8 bit - use “Tiff -16 bit” for 12 bit acquired images (Most other software will not recognize 12 bit)

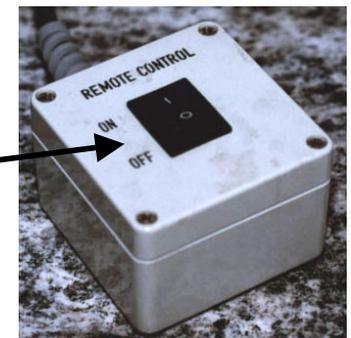
## Shut Down Procedure

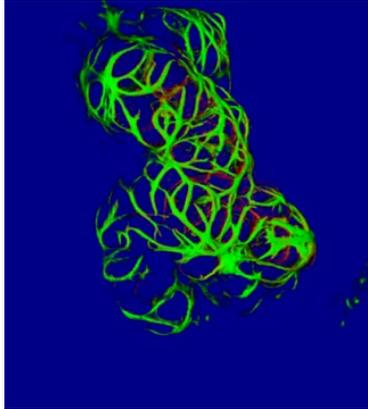
1. Go to: *Acquire* in the LSM menu - *Laser* – and deactivate HeNe Lasers by clicking *Off* to switch off Lasers



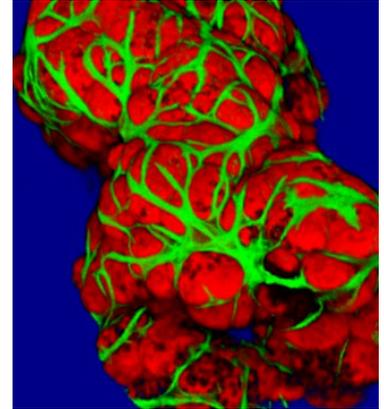
**Note:** To turn off the Argon laser, first click on *Standby*, then reduce output power to 25%. Select *Off*.

2. Go to *File - Exit* to leave LSM 510 program
3. Shut down the computer
4. Wait until fan of Argon laser has switched off.
5. Turn off the remote control box
6. Switch off the mercury vapour lamp.

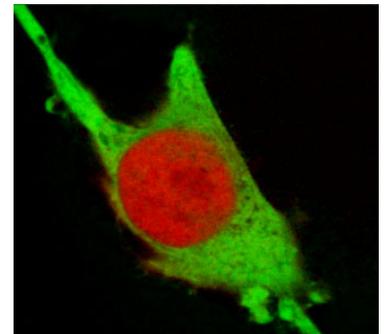
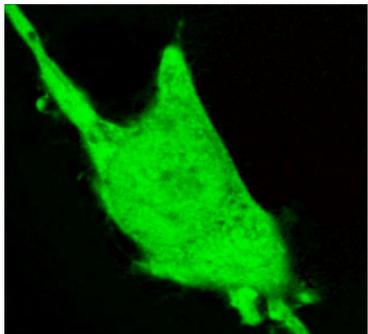




Please note:  
This guided tour is intended merely as a quick introduction into the Zeiss LSM 510 software and does not cover all aspects of the system.



**Please consult the manual  
for detailed instructions!**





**This guided tour is based on  
work done by**

**Peter Jordan  
ICRF  
London  
United Kingdom**

**edited and complemented by  
Eva Simbürger, Solveig Hehl and René Hessling  
Carl Zeiss Jena GmbH**